

Highly Active Antiretroviral Therapy and Anti-tuberculosis Drug Interactions with Associated Clinical Implications: A Review

Karanja JK^{1*}, Kiboi NG², Nebere SN² and Achieng HO¹

¹Department of Zoological Sciences, School of Pure and Applied Sciences, Kenyatta University, Nairobi, Kenya

²Department of Biochemistry and Biotechnology, School of Pure and Applied Sciences, Kenyatta University, Nairobi, Kenya

*Corresponding author: Karanja JK, Department of Zoological Sciences, School of Pure and Applied Sciences, Kenyatta University, Nairobi, Kenya, Tel: +254726721424; E-mail: jkaranja43@yahoo.com

Received date: April 22, 2016; Accepted date: May 06, 2016; Published date: May 20, 2016

Copyright: © 2016 Karanja JK, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Tuberculosis is the most common opportunistic infection associated with HIV/AIDS, and remains a disease of global significance. Co-infection with HIV complicates proper TB diagnosis and therapeutic outcomes. Profound immunosuppression characterizes HIV/TB co-infection prompting early initiation of HAART during TB treatment. Effective management of the co-infection requires concomitant administration of ART and anti-tuberculosis drugs; however, this therapeutic approach has had its fair share of challenges including: overlapping drug toxicities, drug-drug interactions and immune reconstitution reactions. For instance, combination of nevirapine-based ART and rifampicin-based TB treatment is reported to cause hepatotoxicity in healthy volunteers. As such, this review compiles information from multiple studies describing drug interactions associated with co-treatments, with a view to improving management of these co-morbidities.

Keywords: Antiretroviral therapy; Co-infection; Drug interactions; Tuberculosis; HIV

Abbreviations

TB: Tuberculosis; HIV: Human Immunodeficiency Virus; HAART: Highly Active Antiretroviral Therapy; NRTIs: Nucleoside Reverse Transcriptase Inhibitors; NNRTIs: Non-Nucleoside Reverse Transcriptase Inhibitors; PIs: Protease Inhibitors; FIs: Fusion Inhibitors; CRAs: Chemokine Receptor Antagonists; AZT: Zidovudine; ABC: Abacavir; NVP: Nevirapine; EFV: Efavirenz; INH: Isoniazid; RMP: Rifampicin; RTV: Ritonavir; CES: Carboxyl Esterase

Introduction

Tuberculosis (TB) and HIV remain the main cause of high infectious disease burden globally. Sub-Saharan Africa is worst hit by the HIV epidemic and accounts for an estimated 24.7 million cases, compared to global reports indicating 35 million people living with HIV (PLWH) (Figure 1) [1]. TB is the most common presenting illness among PLWH, with both co-morbidities leading to increased morbidity and mortality worldwide [2]. In 2013, global prevalence of HIV/TB stood at 14 million, with 9 million new cases and 1.5 million reported deaths during the same year (Figure 2) [3].

Human Immunodeficiency Virus (HIV) is a major confounder to proper diagnosis and management of TB [4]. This has necessitated the development of highly sensitive tests, including but not limited to; culture systems and nucleic acid amplification assays that are superior to sputum smear microscopy [5]. Interestingly, patients presenting with HIV are at higher risk of developing Multi-Drug Resistant TB (MDR-TB), arguably due to high pill burden, undesired side effects and poor adherence [6].

Currently, there is no defined drug for the cure of HIV infection. Combined Antiretroviral Therapy (cART), also known as Highly Active Antiretroviral Therapy (HAART) is a regimen that merges at least three antiretroviral drugs from different classes of ART in the treatment of HIV [7]. At least six categories of ART exist including; Protease Inhibitors (PI's), Nucleoside and Nucleotide Reverse Transcriptase Inhibitors (NRTI's, NtRTTI's), Non-Nucleoside Reverse Transcriptase Inhibitors (NNRTI's), Fusion Inhibitors (FI's), Integrase Strand Transfer Inhibitors (INSTI's), and Chemokine Receptor Antagonists (CRAs) [8]. Like most other treatment agents HAART is associated with a number of adverse effects including; hepatotoxicity, hypersensitivity rash, lactic acid, osteoporosis, lypodystrophy and metabolic complications (Table 1) [9].

| A) RMP-based TB regimen | | | |
|-------------------------|------------------|----------------------|--|
| | Recommended dose | Nucleoside backbone* | |
| NNRTI | | | |
| EFV | 600 or 800 mg/qd | 2 NRTI/NtRTIs | |
| NVP | 200 mg bid | 2 NRTI/NtRTIs | |

| | | | |
|--|---|----------------------|------------------------------|
| PI | | | |
| SQV/r | 400/400 mg bid or 1000/100 mg bid | 2 NRTI/NtRTIs | |
| LPV/r | 400/400 mg bid | | |
| | | | |
| B) RBT-based TB regimen | | | |
| | Recommended dose | Nucleoside backbone* | Recommended RBT dose |
| PI or NNRTI | | | |
| IDV | 1000 mg tid | 2 NRTI/NtRTIs | 150 mg qd or 300 mg 3×/week |
| NFV | 1000 mg tid | 2 NRTI/NtRTIs | 150 mg qd or 300 mg 3×/week |
| APV | 1200 mg bid | 2 NRTI/NtRTIs | 150 mg qd or 300 mg 3×/week |
| ATV | 400 mg qd | 2 NRTI/NtRTIs | 150 mg qd or 300 mg 3×/week |
| LPV/r | 400/100 mg bid | 2 NRTI/NtRTIs | 150 mg qd or 300 mg 3×/week |
| FPV | 1040 mg bid | 2 NRTI/NtRTIs | 150 mg qd or 300 mg 3×/week |
| RTV combined with ATV, APV, IDV, FPV, SQV | | 2 NRTI/NtRTIs | 150 mg qod or 150 mg 3×/week |
| NVP | 200 mg bid | 2 NRTI/NtRTIs | 300 mg qd or 300 mg 3×/week |
| EFV | 600 mg qd | 2 NRTI/NtRTIs | 600 mg qd or 600 mg qod |
| | | | |
| C) Non-rifamycin-based TB regimen | | | |
| | Usual dose | Nucleoside backbone* | |
| PI | | | |
| IDV | 800 mg tid | 2 NRTI/NtRTIs | |
| NFV | 1250 mg bid or 750 mg tid | 2 NRTI/NtRTIs | |
| APV | 1200 mg bid | 2 NRTI/NtRTIs | |
| ATV | 400 mg qd | 2 NRTI/NtRTIs | |
| LPV/r | 400/100 mg bid | 2 NRTI/NtRTIs | |
| FPV | 1400 mg bid | 2 NRTI/NtRTIs | |
| SQV (soft gel capsule) | 1200 mg tid | 2 NRTI/NtRTIs | |
| RTV boosted PI | | | |
| ATV/r | 300/100 mg bid | 2 NTRI/NtRTIs | |
| AMP/r | 600/100 mg bid or 1200/200 mg qd | 2 NTRI/NtRTIs | |
| IDV/r | 400/400 or 800/100 or 800/200 mg bid | 2 NTRI/NtRTIs | |
| FPV/r | 700/100 mg bid or 1400/200 mg qd | 2 NTRI/NtRTIs | |
| SQV/r | 400/400 mg or 1000/100 bid or 1600/200 qd | 2 NTRI/NtRTIs | |
| NNRTI | | | |
| NVP | 200 mg bid | 2 NTRI/NtRTIs | |

| | | | |
|-----|------------|---------------|--|
| EFV | 600 mg bid | 2 NTRI/NNRTIs | |
|-----|------------|---------------|--|

Table 1: Recommended doses of antiretroviral and RBT-based regimens that can be co-administered in HIV-1 and TB combined therapy.

Multivariate drug factors and HIV/TB co-infections have confounded the management of both diseases resulting in various clinical implications including; liver impairment, renal failure and cardiovascular disorders. Therefore, this review summarizes data on HAART and anti-tuberculous drug interactions, adverse reactions and host toxicity.

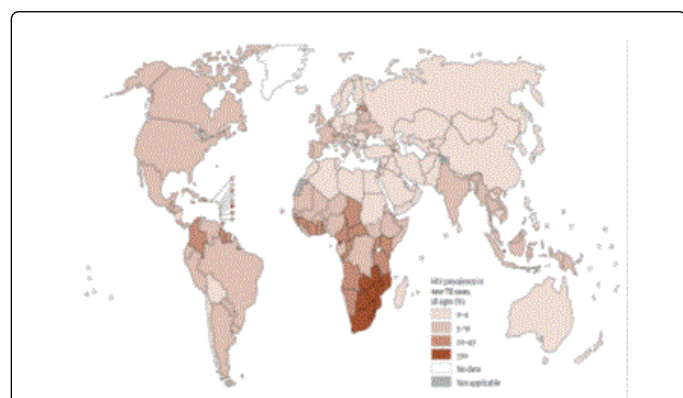


Figure 1: Estimated HIV prevalence in new and relapse TB cases, 2014 [10].

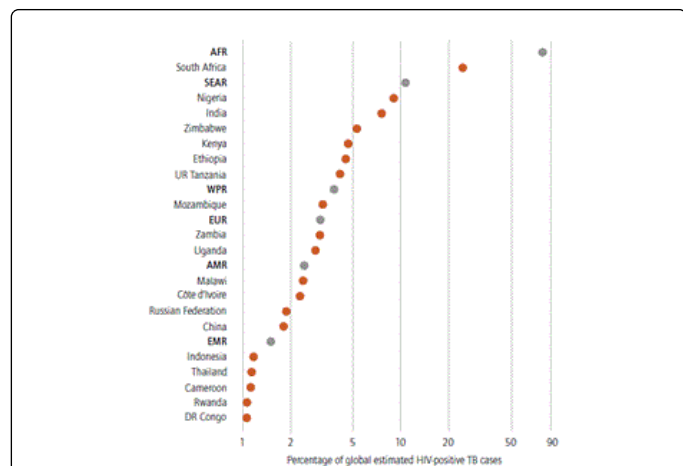


Figure 2: Geographic distribution of the estimated number of human immunodeficiency virus (HIV)-positive tuberculosis cases. For each country (red circles) and World Health Organization region (gray circles), the number of incident tuberculosis cases arising in people infected with HIV is shown as a percentage of the global total of such cases. Note: AFR, African region; AMR, American region; DR Congo, Democratic Republic of the Congo; EMR, Eastern Mediterranean region; EUR, European region; SEAR, Southeast Asian region; TB, tuberculosis; UR Tanzania, United Republic of Tanzania; WPR, Western Pacific region [11].

Metabolism of Antiretroviral Drugs

Nucleoside reverse transcriptase inhibitors constitute the backbone of ART regimens. Some commonly used NRTIs including Abacavir (ABC) and Zidovudine (AZT) are metabolized via hepatic glucuronidation and are phosphorylated into their active triphosphate form [12,13]. These drugs are substrates for phase II metabolizing enzymes that do not involve the CYP450 system, thus, they are less prone to interactions with CYP450 substrates such as isoniazid [14].

Non-nucleoside reverse transcriptase inhibitors are usually co-administered with NRTIs for HIV treatment in resource-limited settings. Unlike NRTIs, NNRTIs are not activated by phosphorylation but are metabolised by the CYP450 system, leading to various drug-drug interactions form [15]. Efavirenz (EFV), an NNRTI is metabolized to inactive hydroxylated metabolites by CYP3A4 and CYP2B6 [16]. Studies have shown EFV to be both an inducer and inhibitor of CYP3A4, thus affects metabolism of many other drugs metabolized by the same isoenzyme [17]. Another NNRTI, Nevirapine (NVP) is eliminated via CYP3A4 and CYP2B6 isoenzymes and induces CYP3A4 [18].

Protease inhibitors are recommended as second-line ARV regimens in some resource-constrained setups. All PIs are extensively metabolized by CYP3A4 isoenzyme, with Ritonavir (RTV) having the most pronounced inhibitory effect and Saquinavir (SQV) the least [19]. The potent CYP3A4 inhibitory properties of RTV have been pharmacologically used to boost the concentrations of other PIs when used in combination [20]. Thus, when used as a booster, RTV acts as a therapeutic enhancer rather than as antiviral agent.

Despite integrase inhibitors being limited in developing countries, this review highlights their metabolism and subsequent drug interactions with anti-TB agents. Enfuvirtide, a synthetic peptide fusion inhibitor is shown to be metabolized by proteolytic hydrolysis without involvement of the CYP450 system, thus is less prone to interactions with CYP450 substrates [21]. On the other hand, Maraviroc (MVC) is a substrate of CYP3A4, and dosage adjustments have been recommended in presence of drugs that alter action of this isoenzyme. For instance, the dosage of maraviroc should be increased if combined with CYP3A4 inducers such as Rifampicin (RMP) [22]. The only well characterized integrase strand transfer inhibitor is Raltegravir (RAL) that is reported to be metabolized by glucuronidation and does not interact with CYP450 enzymes [23]. As such, RAL is expected to have minimal drug-drug interactions. However, recent studies indicate potential drug-drug interactions with strong CYP450 inducers such as RMP [24]. Therefore, raltegravir is recommended not to be co-administered with RMP since it lowers raltegravir plasma concentrations.

Metabolism of anti-tuberculosis drugs

First-line anti-tubercular drugs include isoniazid, rifampicin, pyrazinamide, ethambutol and streptomycin, with isoniazid and rifampicin identified as the most active agents [3]. Following instances of resistance to first-line agents and serious drug reactions, second line

drugs including fluoroquinolones, cycloserine and kanamycin may be administered [11,25].

Isoniazid (INH), a highly potent anti-tubercular agent, undergoes metabolism in the liver by acetylation through the genetically polymorphic N-acetyltransferase 2 (NAT2) enzyme. INH is metabolized internally to acetylisoniazid, and then undergoes hydrolysis to isonicotinic acid and acetylhydrazine [26,27]. The drug is a substrate for phase II metabolic enzymes and does not interact with CYP450 system, thus, is not prone to cross-reactions with the CYP450 substrates [28].

Rifampicin, an anti-TB drug is metabolized by human carboxyl esterase (CES) via deacetylation within liver microsomes [29]. Hence, the drug is an N-acetyltransferase inhibitor that causes decreased acetylation ratio in fast acetylators. Additionally, both RMP and rifabutin are known potential inhibitors of β -subunit-dependent DNA-RNA polymerase which limits DNA formation by *M. tuberculosis* [30]. As a strong inducer of most CYP450 isoforms including CYPs 1A2, 2C9, 2C19, 2D6, and 3A4, RMP is reported to hasten elimination of many drugs such as protease inhibitors and some NNRTIs that are also substrates of CYP450 enzymes [20,31].

Pyrazinamide and ethambutol also make up essential medications in TB therapy. The former is primarily eliminated by hepatic metabolism and involves two pathways that differ by the order of succession of enzymatic sequences but yields similar end products comprising 5-hydroxypyrazinoic acid and pyrazinoic acid [32]. Antimicrobial potency of pyrazinamide has been suggested to be mediated through its conversion to pyrazinoic acid by the amidase activity of intracellular tubercle bacilli and subsequent entrapment in phagosomes [33]. On the other hand, ethambutol is poorly metabolized and upto 80% undergoes renal clearance [34], but during instances of renal insufficiency it may accumulate in patients thus heightening nephrotoxicity [35].

Potential Clinical Risks of Drug Interactions

Drug-drug interactions

Most clinically important drug-drug interactions occur during metabolism of drugs. Numerous phase I metabolic processes take place in the hepatic microsomes via Cytochrome P450 (CYP450) family of heme-containing mono-oxygenases [36]. Previous reports indicate that drugs inducing or inhibiting CYP450 enzymes may either decrease or increase concentrations of concurrently administered drugs [37]. Therefore, changes in drug concentrations resulting from drug interactions may bring about treatment failure or toxicities.

For instance, RMP-based anti-tuberculous therapy induces multiple genes that control drug metabolism and transport including; cytochrome P450 isoenzymes and the drug efflux pump p-glycoprotein [38]. Thus, RMP has the potential to reduce plasma concentrations of concomitantly administered antiretroviral agents that eventually results in inadequate plasma levels and poor ART outcomes. Previous studies documented marked reduction in EFV concentrations following RMP-based TB therapy due to induction of CYP2B6 and CYP3A5 isoenzymes [39,40]. Similarly, plasma NVP levels decline significantly following concomitant use with RMP in treatment of HIV-1 and TB co-infection [41]. Hence co-administration of NVP/EFV and RMP during combined therapy requires utmost consideration in order to avoid lowering treatment efficacy.

Efavirenz, a Non-Nucleoside Reverse Transcriptase Inhibitor (NNRTI) that is also used alongside TB therapy has been implicated with teratogenic effects during pregnancy [42]. As a result, EFV is substituted with alternative regimen if available, especially in resource-constrained settings. On the other hand, cytochrome P450 2B6 516 G>T gene polymorphism exhibits dominance among Africans and impairs metabolism of EFV thereby maintaining high concentrations at EFV standard doses even during co-treatment of TB [41,43].

Nevirapine, a potent NNRTI and RMP, a first line anti-tubercular drug are both used in HIV and TB co-infected patients [44]. However, concurrent use of both therapeutic agents is not recommended because RMP is a potent inducer of hepatic CYP450, which in turn, interferes with metabolism of NVP [45]. Prior studies suggest that oxidative metabolism of NVP is mediated primarily by CYP isozymes from the CYP3A4 family [46]. By inducing the expression of CYP3A4 isoenzyme in the liver, RMP greatly reduces the plasma concentration of NVP upon concurrent administration [45].

Protease inhibitor-based antiretroviral regimens such as Lopinavir-Ritonavir (LPV/r) and Darunavir (DRV) are important option for the treatment of HIV infection [47]. However, studies have demonstrated that co-administration of PIs with RMP reduces PIs systemic concentration to less than 75% thereby compromising HIV treatment efficacy [48]. In order to evaluate how to boost the PIs plasma concentrations when concurrently administered with RMP, several studies have been conducted to assess either higher doses of the PI or of the pharmacologic boosting agent, RTV, or both [48,49]. These studies indicate that PIs plasma concentrations could be boosted by two methods; super-boosting, (administering PI with higher dose of RTV) and double dosing, (doubling the dose of both the PI and RTV). Although these strategies may result in adequate protease inhibitor concentrations, clinical reports have documented increased hepatotoxicity [49,50].

Among the rifamycins, the drug RMP has earlier been shown to be the most powerful inducer of CYP3A4 [51], hence responsible for clinically important interactions with PIs and NNRTIs. However, other than RMP, the anti-tubercular agent RBT also induces CYP3A4 isoenzyme but to a lesser magnitude [52]. Interestingly though, RBT is also a substrate of the enzyme unlike RMP [53]. As such, inhibitors of CYP3A4 including PIs and NNRTIs will essentially elevate plasma concentrations of RBT, with no effect on RMP metabolism. For instance, concomitant administration of RBT with NVP (NNRTI) or RTV (PI) results in elevated systemic levels of RBT [54]. Thus dose adjustments of RBT are required in order to control toxicity [55].

Antiretroviral backbone regimen that comprises of NNRTIs including; AZT, TDF, 3TC, ABC and ddI among others, have been described not to elicit major clinically significant drug interactions with various anti-TB regimen specifically RBT and the commonly used RMP [12]. However, previous studies conducted by Burger et al. documented notable drug interactions between RMP and AZT. The continuous administration of both AZT and RMP regimens concurrently, lead to marked clearance of plasma AZT levels with subsequent therapeutic implications [56]. This activity may result from RMP CYP450 powerful inducing capacity. Contrastingly, other reports document AZT substantially lowering systemic levels of pyrazinamide, also an anti-tuberculosis agent [57], which may be owed to the fact that pyrazinamide is a less potent inducer of CYP450 as compared to RMP.

Complex toxic effects

Toxicity profiles of antiretrovirals and anti-tuberculosis drugs overlap making it complex to identify the exact causative agent [58]. More importantly, concomitant administration of NNRTIs and boosted PIs, with TB treatment has been shown to accelerate drug induced liver injury (DILI), which may heighten drug resistance and ultimate treatment failure [59-61]. Similarly, co-administration of aminoglycosides such as: kanamycin and amikacin used for drug-resistant TB, and tenofovir (TFV) an NRTI aggravates nephrotoxicity [62,63].

Among the identified predictors of anti-tubercular and antiretroviral associated DILI include; slow acetylation status, increased baseline liver aminotransferases, reduced haemoglobin and albumin levels, marked elevation of plasma efavirenz concentration and also *CYP2B6*6/*6* and *ABC13435TT* genotypic characterization [59]. On the whole, impaired liver functions greatly complicate the management of the co-epidemic and may necessitate withdrawal of hepatotoxic antiretrovirals and TB drugs [64], a clinical practice which though necessary in case of severe toxicities, tends to worsen prognosis.

Shared adverse drug effects

Adverse drug reactions resulting from concurrent treatment of HIV and TB are common among the dual infections, and predispose patients mainly to liver damage due to shared metabolic pathways [65,66]. A high incidence of peripheral neuropathy (55%) has been documented in patients undergoing both d4T and INH treatment [67], that may be as a consequence of additive toxic effects from both therapeutic agents. To add further, individuals on INH treatment should closely be monitored and require administration of supplemental pyridoxine therapy in order to minimize risk of INH-related CNS/neurotoxicity [68]. On the other hand, concomitant use of both NVP and anti-TB drugs especially RMP subjects multiple overlapping toxicities including hypersensitivity skin rash and hepatitis [44,69,70].

The risk of hepatotoxicity is up-regulated during antiretroviral and anti-TB therapy, hence the need to screen for pre-existing liver diseases including; hepatitis B and C before HAART or anti-TB commencement [71]. In individuals exhibiting abnormal baseline hepatic transaminases, an elevation of two-to three fold above abnormal baseline levels should be adopted as threshold for

hepatotoxicity [72]. On the other hand, AZT administration has been discouraged in patients with low haemoglobin levels (<8 g/dl) due to likelihood of developing AZT associated anaemia [73]. The antiretroviral drug is also implicated with inducing myelosuppression in HIV positive patients [74]. Finally, gastrointestinal disturbances including malabsorption are reported with all first line anti-TB drugs and various antiretroviral regimen including NVP, that may possibly be attributed to presence of gastrointestinal disease (Tables 2 and 3) [75].

| Toxicity/side effect | Antiretroviral drugs | Anti-tuberculosis drugs |
|-------------------------|-----------------------------|--|
| Skin rash | ABC, NVP, EFV, APV, FPV | INH, RMP, pyrazinamide, quinolones |
| Peripheral neuropathy | d4T, ddl, ddC | INH, cycloserine, ethambutol |
| CNS toxicity | EFV | INH, streptomycin, quinolones, cycloserine |
| Hepatotoxicity | EFV, NVP, all PIs and NRTIs | RMP, RBT, INH, pyrazinamide |
| Anaemia, neutropenia | AZT | RMP, INH |
| Bone marrow suppression | AZT | RBT, RMP |
| Ocular effects | ddl, | RBT, ethambutol |
| Nausea, vomiting | RTV, IDV, AZT | RMP, quinolones, ethionamide, pyrazinamide |
| GIT side effects | All | All |
| Hepatitis | NVP, PIs | RMP, INH, ethionamide, pyrazinamide |

Note: NVP: Nevirapine; EFV: Efavirenz; ABC: Abacavir; APV: Amprenavir; FPV: Fosamprenavir; d4T: Stavudine; ddl: Didanosine; ddC: Zalcitabine; AZT: Zidovudine; INH: Isoniazid; RMP: Rifampicin; RBT: Rifabutin; PIs: Protease inhibitors; NRTIs: Nucleoside reverse transcriptase inhibitors; GIT: Gastrointestinal tract [44,70].

Table 2: Overlapping or additive adverse effect profiles due to antiretroviral and anti-tuberculosis agents.

| Event/issue | Management suggestion (s) |
|---|---|
| Overlapping side effect profiles of antiretrovirals and anti-tuberculosis drugs | Defer ART until there has been time to identify and manage side effects from anti-tuberculosis drugs (1-2 mo) |
| Drug interactions between rifamycins and antiretroviral agents (HIV-1 PIs and NNRTIs) | Use RBT with recommended dose adjustments in Table 1. Use RMP with EFV and RTV (at doses of >400 mg bid) |
| Paradoxical reactions after initiating antiretroviral treatment | Delay ART until after TB treatment if CD4 cell count is relatively high (>300µL) In patients with low CD4 cell counts, defer ART until TB is substantially improved (2 mo) Schedule clinical follow-up soon after onset of ART to detect paradoxical reactions and/or drug side effects early |

Note: HIV-1: Human immune deficiency virus type 1; TB: Tuberculosis; ART: Antiretroviral therapy; PIs: Protease inhibitors; RBT: Rifabutin; RMP: Rifampicin; EFV: Efavirenz; RTV: Ritonavir; mg: Milligram; bid: Twice daily; mo: Months; µL: Microlitre [76].

Table 3: Summarized management recommendations for ART use in HIV and TB co-infected patients.

Tuberculosis Immune Reconstitution Inflammatory Syndrome (TB-IRIS)

Immune Reconstitution Inflammatory Syndrome (IRIS) is the transient deterioration of signs and symptoms of tuberculosis after initiation of ART, despite reduction in HIV viral load and immunological recovery [77,78]. Two forms of IRIS exist: Paradoxical TB-IRIS which occurs in patients diagnosed with TB and already established on TB treatment prior to ART, and they present with recurrent or new TB; unmasking TB-IRIS occurring in patients not on TB treatment when they initiate ART, a form characterized by an unusually high inflammatory response of TB [78,79]. Pronounced IRIS features comprise: recurrent TB symptoms, lymph node enlargement, fever, cold abscess, worsening respiratory signs and central nervous system lesions [77]. Abdominal manifestations have also been reported and include intestinal lesions, splenic and hepatic derangements, peritonitis, ascites as well as lymphadenopathy [77,80]. Hepatic features frequently occur in about 21-56% of TB-IRIS patients, and is usually difficult to differentiate with drug-induced hepatitis. Major clinical features are liver enlargement, liver functional derangements and granulomatous hepatitis [81,82].

New cases of paradoxical TB-IRIS account for 8-43% among patients who initiate ART while on TB therapy. Key risk factors for this

condition are low CD4⁺ T cell counts, disseminated TB and short interval between starting TB treatment and ART [83,84]. Contrastingly, a study carried out among Ugandan patients found no significant association between interval of starting treatment of the dual infections and the development of TB-IRIS. The study reported that delaying ART until 2 months of TB treatment did not appear to deter paradoxical TB-IRIS [85]. These observed variations may result in distinct mechanisms of immune activation that are differentially affected by antiretroviral treatment. On the other hand, fatalities associated with paradoxical TB-IRIS are rare, only exceptionally reported in cases where central nervous system is affected [86].

Diagnosis of paradoxical TB-IRIS is complicated by the lack of confirmatory diagnostic tests [87]. Opportunistic infections, malignancies and drug resistance have to be excluded during assessment. However, in resource-challenged settings, a diagnosis of TB-IRIS can also be performed based on case definition as recommended by the International Network for the Study of HIV-associated IRIS (INSHI) [78]. On the other hand, management of paradoxical TB-IRIS can be done using non-steroidal anti-inflammatory drugs (NSAIDs) and steroids [83] (Table 4).

| Study no. | Study, year | Years studied | Incidence, proportion, % | Median Age of patients, years | Median CD4 count, cells/µL | Median Viral load, copies/mL | Median time, days from TB diagnosis and treatment to IRIS | Median time, days from ART start to development of IRIS |
|-----------|----------------------------|---------------|--------------------------|-------------------------------|----------------------------|------------------------------|---|---|
| 1. | Narita et al. 1998 [5] | 1996-1997 | 12/33 (36) | 40 ^a | 51 ^a | 5.80 | 109 ^a | 15 ^a |
| 2. | Breton et al. 2004 [6] | 1996-2001 | 16/37 (43) | 35 | 100 | 5.36 | 48 | 12 |
| 3. | Breen et al. 2004 [7] | | 14/50 (28) | 36 | N/A | N/A | 33 | 11 |
| 4. | Kumarasamy et al. 2004 [8] | 2000-2003 | 11/144 (8) | 29 | 123 | N/A | 42 | 22 |
| 5. | Lawn et al. 2007 [9] | 2002-2005 | 19/160 (12) | 35 | 68 | 4.84 | 105 | 14 |

Note: ART: Antiretroviral therapy; NA: Not available; TB: Tuberculosis; HIV: Human immunodeficiency virus; ^aMean.

Table 4: Incidences of tuberculosis-immune reconstitution inflammatory syndrome (IRIS) in HIV-TB co-infection.

Conclusions and Future Directions

This review compiles data from various sources on multiple adverse drug effects stemming from concomitant use of ART and anti-tuberculosis drugs. These include; d4T and INH-induced peripheral neuropathy, NVP and RMP associated hypersensitivity rash and AZT induced myelosuppression among others. However, these adverse effects require to be ascertained through appropriate clinical examination for specific signs and symptoms that will eventually aid in improved patient management. Equally, although ART and anti-tuberculosis regimen improves patient outcomes, several drug-drug

interactions have been documented during the concurrent use of both therapeutic agents. Nonetheless, these interactions subject patients to overlapping toxicities with associated clinical implications. As such, more investigations on actual pharmacokinetic mechanisms behind drug interaction is necessitated, while also factoring in patient safety and treatment efficacy. Several clinical trials may generate answers to these concerns.

Authors' Contributions

All authors contributed in drafting, review of article and revising the manuscript. Final version of the manuscript was approved by all authors.

Acknowledgment

The authors thank Kenyatta University library for facilitating space and accessibility to online databases used to gather relevant information for this review. This work received no specific grant from any funding agency whatsoever.

References

- (2014) UNAIDS, JUNP on H. The gap report. Geneva.
- WHO (2011) Guidel Intensified Tuberc Case-Find Isoniazid Prev Ther People Living HIV Resour-Constrained Settings. Switzerland.
- Zumla A, George A, Sharma V, Herbert RHN, Ilton BM, et al. (2015) The WHO 2014 global tuberculosis report—further to go. *Lancet Glob Health* 3: e10-e12.
- Mendelson M (2007) Diagnosing tuberculosis in HIV-infected patients: challenges and future prospects. *Br Med Bull* 81: 149-165.
- Swaminathan S, Padmapriyadarsini C, Narendran G (2010) HIV-associated tuberculosis: clinical update. *Clin Infect Dis* 50: 1377-1386.
- Kumwenda JJ (2014) TB/HIV facts 2012-2013.
- Moyle GJ, Gazzard BG, Cooper DA, Gatell J (1998) Antiretroviral therapy for HIV infection. *Drugs* 55: 383-404.
- Murphy EL, Collier AC, Kalish LA, Assmann SF, Para MF, et al. (2001) Highly active antiretroviral therapy decreases mortality and morbidity in patients with advanced HIV disease. *Ann Intern Med* 135: 17-26.
- Subbaraman R, Chaguturu SK, Mayer KH, Flanigan TP, Kumarasamy N, et al. (2007) Adverse effects of highly active antiretroviral therapy in developing countries. *Clin Infect Dis* 45: 1093-1101.
- WHO (2012) WHO policy on collaborative TB/HIV activities: guidelines for national programmes and other stakeholders. Geneva.
- (2010) WHO. Global tuberculosis control.
- Pozniak AL, Coyne KM, Miller RF, Lipman MCI, Freedman AR, et al. (2011) British HIV Association guidelines for the treatment of TB/HIV coinfection 2011. *HIV Med* 12: 517-524.
- Katzung BG, Masters SB, Trevor AJ (2009) Basic and Clinical Pharmacology. McGraw-Hill Companies, New York, USA.
- Dooley KE, Charles F, Andrade AS (2008) Drug interactions involving combination antiretroviral therapy and other anti-infective agents: repercussions for resource-limited countries. *J Infect Dis* 198: 948-961.
- Joly V, Yeni P (2000) [Non-nucleoside reverse transcriptase inhibitors]. In: *Annales de medecine interne* pp: 260-267.
- Bumpus NN, Kent UM, Hollenberg PF (2006) Metabolism of efavirenz and 8-hydroxyefavirenz by P450 2B6 leads to inactivation by two distinct mechanisms. *J Pharmacol Exp Ther* 318: 345-351.
- Hariparsad N, Nallani SC, Sane RS, Buckley DJ, Buckley AR, et al. (2004) Induction of CYP3A4 by efavirenz in primary human hepatocytes: comparison with rifampin and phenobarbital. *J Clin Pharmacol* 44: 1273-1281.
- Smith PF, DiCenzo R, Morse GD (2001) Clinical pharmacokinetics of non-nucleoside reverse transcriptase inhibitors. *Clin Pharmacokinet* 40: 893-905.
- Koudriakova T, Iatsimirskaia E, Utkin I, Gangl E, Vouros P, et al. (1998) Metabolism of the human immunodeficiency virus protease inhibitors indinavir and zalcitabine by human intestinal microsomes and expressed cytochrome P4503A4/3A5: mechanism-based inactivation of cytochrome P4503A by zalcitabine. *Drug Metab Dispos* 26: 552-561.
- Baciewicz AM, Chrisman CR, Finch CK, Self TH (2013) Update on rifampin, rifabutin, and rifapentine drug interactions. *Curr Med Res Opin* 29: 1-12.
- Patel IH, Zhang X, Nieforth K, Salgo M, Buss N (2005) Pharmacokinetics, pharmacodynamics and drug interaction potential of enfuvirtide. *Clin Pharmacokinet* 44: 175-186.
- Mannu J, Jenardhanan P, Mathur PP (2011) A computational study of CYP3A4 mediated drug interaction profiles for anti-HIV drugs. *J Mol Model* 17: 1847-1854.
- Kassahun K, McIntosh I, Cui D, Hreniuk D, Merschman S, et al. (2007) Metabolism and disposition in humans of raltegravir (MK-0518), an anti-AIDS drug targeting the human immunodeficiency virus 1 integrase enzyme. *Drug Metab Dispos* 35: 1657-1663.
- Wenning LA, Hanley WD, Brainard DM, Petry AS, Ghosh K, et al. (2009) Effect of rifampin, a potent inducer of drug-metabolizing enzymes, on the pharmacokinetics of raltegravir. *Antimicrob Agents Chemother* 53: 2852-2856.
- Crofton JW, Choulet P, Maher D (1997) Guidelines for the management of drug-resistant tuberculosis. World Health Organization.
- Zabost A, Brzezińska S, Kozińska M, Błachnio M, Jagodziński J, et al. (2013) Correlation of N-acetyltransferase 2 genotype with isoniazid acetylation in Polish tuberculosis patients. *Bio Med Res Int* 2013.
- Preziosi P (2007) Isoniazid: metabolic aspects and toxicological correlates. *Curr Drug Metab* 8: 839-851.
- Guengerich FP (2006) Cytochrome P450s and other enzymes in drug metabolism and toxicity. *AAPS J* 8: E101-E111.
- Jamis-Dow CA, Katki AG, Collins JM, Klecker RW (1997) Rifampin and rifabutin and their metabolism by human liver esterases. *Xenobiotica Fate Foreign Compd Biol Syst* 27: 1015-1024.
- Senol G, Erbaycu A, Özsöz A (2005) Incidence of Cross Resistance Between Rifampicin and Rifabutin in Mycobacterium tuberculosis Strains in Izmir, Turkey. *J Chemother* 17: 380-384.
- Finch CK, Chrisman CR, Baciewicz AM, Self TH (2002) Rifampin and rifabutin drug interactions: an update. *Arch Intern Med* 162: 985-992.
- Lacroix C, Tranvouez JL, Phan HT, Duwoos H, Lafont O (1990) Pharmacokinetics of pyrazinamide and its metabolites in patients with hepatic cirrhotic insufficiency. *Arzneimittelforschung* 40: 76-79.
- Salfinger M, Crowle AJ, Reller LB (1990) Pyrazinamide and pyrazinoic acid activity against tubercle bacilli in cultured human macrophages and in the BACTEC system. *J Infect Dis* 162: 201-207.
- Malone RS, Fish DN, Spiegel DM, Childs JM, Peloquin CA (1999) The effect of hemodialysis on isoniazid, rifampin, pyrazinamide, and ethambutol. *Am J Respir Crit Care Med* 159: 1580-1584.
- Launay-Vacher V, Izzedine H, Deray G (2005) Pharmacokinetic considerations in the treatment of tuberculosis in patients with renal failure. *Clin Pharmacokinet* 44: 221-235.
- Gibson GG, Skett P (2001) Introduction to drug metabolism. Nelson Thornes.
- Lin JH, Lu AY (1998) Inhibition and induction of cytochrome P450 and the clinical implications. *Clin Pharmacokinet* 35: 361-390.
- Niemi M, Backman JT, Fromm MF, Neuvonen PJ, Kivistö KT (2003) Pharmacokinetic interactions with rifampicin. *Clin Pharmacokinet* 42: 819-850.
- Faucette SR, Wang H, Hamilton GA, Jolley SL, Gilbert D, et al. (2004) Regulation of CYP2B6 in primary human hepatocytes by prototypical inducers. *Drug Metab Dispos* 32: 348-358.
- Faucette SR, Zhang TC, Moore R, Sueyoshi T, Omiecinski CJ, et al. (2007) Relative activation of human pregnane X receptor versus constitutive androstane receptor defines distinct classes of CYP2B6 and CYP3A4 inducers. *J Pharmacol Exp Ther* 320: 72-80.
- Cohen K, Grant A, Dandara C, McIlleron H, Pemba L, et al. (2008) Effect of rifampicin-based antitubercular therapy and the cytochrome P450 2B6 516G>T polymorphism on efavirenz concentrations in adults in South Africa. *Antivir Ther* 14: 687-695.

42. Chersich MF, Urban ME, Venter FW, Wessels T, Krause A, et al. (2006) Efavirenz use during pregnancy and for women of child-bearing potential. *AIDS Res Ther* 3: 1.
43. Ramachandran G, Kumar AH, Rajasekaran S, Kumar P, Ramesh K, et al. (2009) CYP2B6 G516T polymorphism but not rifampin coadministration influences steady-state pharmacokinetics of efavirenz in human immunodeficiency virus-infected patients in South India. *Antimicrob Agents Chemother* 53: 863-868.
44. Breen RA, Miller RF, Gorsuch T, Smith CJ, Schwenk A, et al. (2006) Adverse events and treatment interruption in tuberculosis patients with and without HIV co-infection. *Thorax* 61: 791-794.
45. Semvua HH, Kibiki GS, Kisanga ER, Boeree MJ, Burger DM, et al. (2015) Pharmacological interactions between rifampicin and antiretroviral drugs: challenges and research priorities for resource-limited settings. *Ther Drug Monit* 37: 22-32.
46. Erickson DA, Mather G, Trager WF, Levy RH, Keirns JJ (1999) Characterization of the in vitro biotransformation of the HIV-1 reverse transcriptase inhibitor nevirapine by human hepatic cytochromes P-450. *Drug Metab Dispos* 27: 1488-1495.
47. Martinez E, Larrousse M, Llibre JM, Gutierrez F, Saumoy M, et al. (2010) Substitution of raltegravir for ritonavir-boosted protease inhibitors in HIV-infected patients: the SPIRAL study. *Aids* 24: 1697-1707.
48. Acosta EP, Kendall MA, Gerber JG, Alston-Smith B, Koletar SL, et al. (2007) Effect of concomitantly administered rifampin on the pharmacokinetics and safety of atazanavir administered twice daily. *Antimicrob Agents Chemother* 51: 3104-3110.
49. Decloedt EH, McIlleron H, Smith P, Merry C, Orrell C, et al. (2011) Pharmacokinetics of lopinavir in HIV-infected adults receiving rifampin with adjusted doses of lopinavir-ritonavir tablets. *Antimicrob Agents Chemother* 55: 3195-3200.
50. Sulkowski MS, Mehta SH, Chaisson RE, Thomas DL, Moore RD, et al. (2004) Hepatotoxicity associated with protease inhibitor-based antiretroviral regimens with or without concurrent ritonavir. *Aids* 18: 2277-2284.
51. Rae JM, Johnson MD, Lippman ME, Flockhart DA (2001) Rifampin is a selective, pleiotropic inducer of drug metabolism genes in human hepatocytes: studies with cDNA and oligonucleotide expression arrays. *J Pharmacol Exp Ther* 299: 849-857.
52. Perucca E, Grimaldi R, Frigo GM, Sardi A, Mönig H, et al. (1988) Comparative effects of rifabutin and rifampicin on hepatic microsomal enzyme activity in normal subjects. *Eur J Clin Pharmacol* 34: 595-599.
53. Li AP, Reith MK, Rasmussen A, Gorski JC, Hall SD, et al. (1997) Primary human hepatocytes as a tool for the evaluation of structure-activity relationship in cytochrome P450 induction potential of xenobiotics: evaluation of rifampin, rifapentine and rifabutin. *Chem Biol Interact* 107: 17-30.
54. Gallicano K, Khaliq Y, Carignan G, Tseng A, Walmsley S, et al. (2001) A pharmacokinetic study of intermittent rifabutin dosing with a combination of zidovudine and zalcitabine in patients infected with human immunodeficiency virus. *Clin Pharmacol Ther* 70: 149-58.
55. Sun E, Heath-Chiozzi M, Cameron DW, Hsu A, Granneman RG, et al. (1996) Concurrent zidovudine and rifabutin increases risk of rifabutin-associated adverse events. In: Program and abstracts of the XI International Conference on AIDS Vancouver, British Columbia.
56. Burger DM, Meenhorst PL, Koks CH, Beijnen JH (1993) Pharmacokinetic interaction between rifampin and zidovudine. *Antimicrob Agents Chemother* 37: 1426-1431.
57. Rajesh R, Vidyasagar S, Varma DM, Krishnadas N (2011) Highly active antiretroviral therapy induced drug-drug interactions in Indian Human Immunodeficiency Virus positive patients. *J Clin Med Res* 3: 68-72.
58. McIlleron H, Meintjes G, Burman WJ, Maartens G (2007) Complications of antiretroviral therapy in patients with tuberculosis: drug interactions, toxicity, and immune reconstitution inflammatory syndrome. *J Infect Dis* 196: S63-S75.
59. Yimer G, Ueda N, Habtewold A, Amogne W, Suda A, et al. (2011) Pharmacogenetic & pharmacokinetic biomarker for efavirenz based ARV and rifampicin based anti-TB drug induced liver injury in TB-HIV infected patients. *PLoS One* 6: e27810.
60. Tostmann A, Boeree MJ, Aarnoutse RE, De Lange W, Van Der Ven AJ, et al. (2008) Antituberculosis drug-induced hepatotoxicity: concise up-to-date review. *J Gastroenterol Hepatol* 23: 192-202.
61. Kontorinis N, Dieterich DT (2003) Toxicity of non-nucleoside analogue reverse transcriptase inhibitors. In: Seminars in liver disease pp: 173-182.
62. Coyne KM, Pozniak AL, Lamorde M, Boffito M (2009) Pharmacology of second-line antituberculosis drugs and potential for interactions with antiretroviral agents. *Aids* 23: 437-446.
63. Harries AD, Zachariah R, Lawn SD (2009) Providing HIV care for co-infected tuberculosis patients: a perspective from sub-Saharan Africa [State of the art series. Tuberculosis. Edited by ID Rusen. Number 3 in the series]. *Int J Tuberc Lung Dis* 13: 6-16.
64. Tansuphasawadikul S, Saito W, Kim J, Phonrat B, Dhitavat J, et al. (2007) Channachanan S, et al. Outcomes in HIV-infected patients on antiretroviral therapy with tuberculosis. *Southeast Asian J Trop Med Public Health* 38: 1053.
65. Yimer G, Aderaye G, Amogne W, Makonnen E, Aklillu E, et al. (2008) Anti-tuberculosis therapy-induced hepatotoxicity among Ethiopian HIV-positive and negative patients. *PLoS One* 3: e1809.
66. Pukenyte E, Lescure FX, Rey D, Rabaud C, Hoen B, et al. (2007) Incidence of and risk factors for severe liver toxicity in HIV-infected patients on anti-tuberculosis treatment. *Int J Tuberc Lung Dis* 11: 78-84.
67. Breen RA, Lipman MC, Johnson MA (2000) Increased incidence of peripheral neuropathy with co-administration of stavudine and isoniazid in HIV-infected individuals. *Aids* 14: 615.
68. Siskind MS, Thienemann D, Kirlin L (1993) Isoniazid-induced neurotoxicity in chronic dialysis patients: report of three cases and a review of the literature. *Nephron* 64: 303-306.
69. Swaminathan S, Luetkemeyer A, Srikantiah P, Lin R, Charlebois E, et al. (2006) Antiretroviral therapy and TB. *Trop Doct* 36: 73-79.
70. Dean GL, Edwards SG, Ives NJ, Matthews G, Fox EF, et al. (2002) Treatment of tuberculosis in HIV-infected persons in the era of highly active antiretroviral therapy. *Aids* 16: 75-83.
71. Yimer G, Gry M, Amogne W, Makonnen E, Habtewold A, et al. (2014) Evaluation of patterns of liver toxicity in patients on antiretroviral and anti-tuberculosis drugs: a prospective four arm observational study in Ethiopian patients. *PLoS One* 9: e94271.
72. Saukkonen JJ, Cohn DL, Jasmer RM, Schenker S, Jereb JA, et al. (2006) An official ATS statement: hepatotoxicity of antituberculosis therapy. *Am J Respir Crit Care Med* 174: 935-952.
73. Agarwal D, Chakravarty J, Chaube L, Rai M, Rani Agrawal N, et al. (2010) High incidence of zidovudine induced anaemia in HIV infected patients in eastern India. *Indian J Med Res* 132: 386.
74. Richman DD, Fischl MA, Grieco MH, Gottlieb MS, Volberding PA, et al. (1987) The toxicity of Azidothymidine (AZT) in the treatment of patients with AIDS and AIDS-related complex. *N Engl J Med* 317: 192-197.
75. Burman WJ, Gallicano K, Peloquin C (1999) Therapeutic implications of drug interactions in the treatment of human immunodeficiency virus-related tuberculosis. *Clin Infect Dis* 28: 419-429.
76. Burman WJ, Jones BE (2001) Treatment of HIV-related tuberculosis in the era of effective antiretroviral therapy. *Am J Respir Crit Care Med* 164: 7-12.
77. Lawn SD, Bekker LG, Miller RF (2005) Immune reconstitution disease associated with mycobacterial infections in HIV-infected individuals receiving antiretrovirals. *Lancet Infect Dis* 5: 361-373.
78. Meintjes G, Lawn SD, Scano F, Maartens G, French MA, et al. (2008) Tuberculosis-associated immune reconstitution inflammatory syndrome: case definitions for use in resource-limited settings. *Lancet Infect Dis* 8: 516-523.
79. Lawn SD, Wilkinson RJ, Lipman MC, Wood R (2008) Immune reconstitution and "unmasking" of tuberculosis during antiretroviral therapy. *Am J Respir Crit Care Med* 177: 680-685.

-
80. Wu SW, Chen CJ, Lin TY, Wang NC (2008) Acute peritonitis as presentations of tuberculosis-associated immune reconstitution inflammatory syndrome in an HIV-infected man. *Am J Med Sci* 335: 387-389.
 81. Lawn SD, Wood R (2007) Hepatic involvement with tuberculosis-associated immune reconstitution disease. *Aids* 21: 2362-2363.
 82. Meintjes G, Rangaka MX, Maartens G, Rebe K, Morroni C, et al. (2009) Novel relationship between tuberculosis immune reconstitution inflammatory syndrome and antitubercular drug resistance. *Clin Infect Dis* 48: 667-676.
 83. Breen RAM, Smith CJ, Bettinson H, Dart S, Bannister B, et al. (2004) Paradoxical reactions during tuberculosis treatment in patients with and without HIV co-infection. *Thorax* 59: 704-707.
 84. Shelburne SA, Visnegarwala F, Darcourt J, Graviss EA, Giordano TP, et al. (2005) Incidence and risk factors for immune reconstitution inflammatory syndrome during highly active antiretroviral therapy. *Aids* 19: 399-406.
 85. Baalwa J, Mayanja-Kizza H, Kamya MR, John L, Kambugu A, et al. (2008) Short Report: Worsening and unmasking of tuberculosis in HIV-1 infected patients after initiating highly active anti-retroviral therapy in Uganda. *Afr Health Sci* 8: 190-195.
 86. Pepper DJ, Marais S, Maartens G, Rebe K, Morroni C, et al. (2009) Neurologic manifestations of paradoxical tuberculosis-associated immune reconstitution inflammatory syndrome: a case series. *Clin Infect Dis* 48: e96-107.
 87. Reyes-Corcho A, Bouza-Jiménez Y (2010) Redefinition of tuberculosis-associated immune reconstitution syndromes in HIV-infected individuals: a need for new clinical tools. *J Infect Dis* 201: 793-794.