

**EFFECTS OF PHOSPHORUS AND SOIL MOISTURE ON GRAIN YIELD, LEAF
AND GRAIN TISSUE CONCENTRATION OF IRON AND ZINC IN THREE BEAN
(*PHASEOLUS VULGARIS L.*) GENOTYPES**

BY

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DECLARATION

Declaration by the Candidate

I HARRISON KATWIKILA MUNUVE declare that this thesis is my original work and has not been presented for the award of a degree in any university or any other award.

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DEDICATION

This research project is dedicated to my wife Peninah Mutethya and children, my late brother James Mutuku for their encouragement during my study period.

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ABSTRACT

Common bean (*Phaseolus vulgaris L.*) is an important source of protein, minerals and food for the majority of the poor population in sub-Saharan Africa. However, its contribution to grain yield and micronutrient level is constrained by moisture stress and low available soil phosphorus. A study was carried out to determine the effects of bean genotypes, P fertilizer and moisture regimes on bean (*P. vulgaris L.*) grain yield and tissue concentration of Zn and Fe in three bean genotypes at KARI, Katumani Research Centre in Machakos, located at 1560 masl. The study was laid out in a randomized complete block design with five P fertilizer treatments in split-split plot arrangement with three replications. The main plot was moisture regimes, subplots were genotypes and the sub-sub plots were P rates. Data on concentration of Fe and Zn at flowering, grain yield and grain concentration of Fe and Zn at bean maturity were recorded. Data for grain yield, grain and leaf tissue concentration of Fe and Zn were analysed using ANOVA, Means were separated using LSD at significant ($P < 0.05$) level and correlation between grain concentration of Fe and Zn, as well as between grain yield and leaf concentration of Fe and Zn were determined. (SAS 8.2; SAS institute, 1999). The results of grain yield showed highly significant ($P = 0.0006$) interaction between moisture regimes, genotypes and P application rates. Grain yield of the three genotypes increased with P application rate up to 60 kg ha^{-1} regardless of the moisture regimes. The leaf and grain concentration of Fe and Zn showed highly significant ($P < 0.01$) interaction between moisture regimes, genotypes and P fertilizer application rate. Similarly, percentage of leaf Fe and Zn accumulated to grain showed highly significant ($P < 0.0001$) interactions between moisture regimes, genotypes and P fertilizer application rate. All genotypes had significantly ($P < 0.05$) higher concentration of Fe and Zn in leaves compared to the grain. Grain Fe concentration was higher in beans grown under adequate moisture conditions than in moisture stressed condition. A highly significant ($n = 90$; $r = 0.79736$; $P < 0.0001$) positive correlation between grain Fe and Zn concentration and highly significant ($n = 90$; $r = 0.53662$; $P < 0.0001$) positive correlation between leaf Fe and Zn concentration were observed. It follows that an increase in grain Fe concentration correlates with an increase in grain Zn concentration, also an increase in leaf Fe concentration correlates with an increase in leaf Zn concentration. A significant ($n = 90$; $r = -0.34860$; $P = 0.0008$) negative correlation between the grain yield and leaf Fe concentration and highly significant ($n = 90$; $r = -0.58292$; $P = 0.0001$) negative correlation between grain yield and leaf Zn concentration were observed. Hence an increase in grain yield correlates with a decrease in leaf concentration of Fe and Zn. In order to increase the grain yield and grain concentration of Fe and Zn, application of P fertilizer, maintaining adequate soil moisture and planting superior bean genotype is required.

Key Words: Phosphorus; Soil Moisture; Yields; Bean; Iron; Zinc.

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LIST OF ABBREVIATION AND ACRONYMS

AEZ	-	Agro Ecological Zone
AGRA	-	Alliance for Green Revolution in Africa
AIDS	-	Acquired Immune Deficiency Syndrome
ANOVA	-	Analysis of Variance
CGIAR	-	Consortium of International Agricultural Research Centers
CCRP	-	Collaborative Crop Research Program
CIAT	-	International Centre for Tropical Agriculture
FAO	-	Food and Agricultural Organisation of the United Nation.
HIV	-	Human Immunodeficiency Virus
ILRI	-	International Livestock Research Institute
KARI	-	Kenya Agricultural Research Institute
LSD	-	Least Significant Difference
Masl	-	Meters above Sea Level
PABRA	-	Pan Africa Bean Research Alliance
TSP	-	Triple Super Phosphate
UK	-	United Kingdom
USA	-	United States of America

RCBD	-	Randomized Complete Block Design
SAS	-	Statistical Analysis System
SE	-	Standard Error
SEKU	-	South Eastern Kenya University
SIDA	-	Swedish International Development Cooperation Agency
V ₁	-	Awash 1
V ₂	-	Awash Melka
V ₃	-	Mexican 142

CHAPTER ONE

INTRODUCTION

1.1 Background Information

Common bean (*Phaseolus vulgaris L.*) is one of the five cultivated species from the genus *Phaseolus* and is a major grain legume crop, third in importance after soya bean and peanut (Broughton *et al.*, 2003). It is one of the principal food and cash crop legume grown in the tropics. It is the primary and least expensive source of calories, protein, dietary fiber, minerals (Fe and Zn) and vitamins for the population in the tropical countries although its intake does not satisfy their mineral requirement (Pachico, 1993; Welch and Graham *et al.*, 1999). Common bean is a small-scale farmer crop in Eastern Africa where it is often cultivated in unfavourable conditions with minimal inputs and is mainly consumed as dried or green beans, the dried beans are sold while raw or as canned beans for local or export market (Katungi *et al.*, 2009). Some of the characteristics that are considered for canning beans include mineral composition, nutrient levels, effective moisture content, percent volume increase, cooking time after soaking, seed dimensions, crude protein, crude fiber, crude fat, carbohydrates and seed colour which is popular in different regions of the world with different cultures (Schoonhoven and Voysest, 1991; Voysest *et al.*, 1994; Gathu, 2012). The growth in demand for pre-cooked beans in the market has necessitated the need to diversify the number of bean varieties on offer thereby creating greater stability of production, increase in nutritional value and food security especially with the looming climate change (Buruchara, 2006).

Bean farming in Kenya is also constrained by decreasing rain due to climatic change leading to reduced grain yields (Remenyik and Nemeske, 2010) and low soil P status accentuated by soil erosion and fixation by oxides in acidic soils (Sixbert and George, 2012). Low soil N and

P levels and acid soil conditions are important constraints for bean production in most of the areas where this crop is grown (Graham *et al.*, 2003). However, in the semi-arid areas where beans are grown the soils are not acidic and are able to fix appreciable amounts of N, if P is supplied (Abdul and Saudi, 2012). This leaves P as a constraint of major concern in the semi-arid areas (Karanja *et al.*, 2011). The decision to fertilizers use in soils with low P levels must be made against the economic background of the cost of fertilizers and the extra crop to be produced (Cooke, 1982). Fertilizer usage has been known to increase grain yield and increase bean nutritional value in terms of mineral contents (McKenzie and Middleton, 1997; Georgina *et al.*, 2007; Sixbert and George, 2012).

Ongoing bean researches addressing the current climate changes have developed heat tolerant beans in Mexico, Nicaragua and Rwanda whose genes are yet to be transferred to bean varieties for breeders in dry land areas (Flora *et al.*, 2011). Studies have shown that an application of P fertilizer led to an increased legume grain yields, particularly with velvet bean, and soya bean (Kamanga *et al.*, 2010). Other studies have shown that bean crop require more P because it is important for nodulation to take place effectively (Ssali and Keya, 1986; Vladimir, 2010). Studies carried out in Columbia showed significant genotype variability on P fertilizer application rates, indicating an increase in Fe concentration in the seeds, but a reduced Zn concentration in seeds (Astudillo *et al.*, 2008).

Studies show varying response of genotypes to shoot biomass, P uptake and yield in a treatment without P which can be utilized in selecting superior genotypes for use in such low soil P environment as breeding material (Sixbert and George, 2012). Selecting improved bean genotypes with high grain yield, high P uptake, moisture stress tolerant and high Zn and Fe concentration for breeding will contribute in addressing the problem of low yields and low dietary mineral intake such as Fe and Zn (Jerome, 2007). The improved breeds can be selected among the genotypes available from various studies that show significant genotype

variability in the tissue concentration of Zn and Fe due to moisture stress reported (Priscila *et al.*, 2008). The aim of this study was to determine the effect of P fertilizer application on grain yield and grain and leaf tissue concentration of Zn and Fe in three bean genotypes grown under different moisture regimes.

1.2 Statement of the Problem

In recent years, bean crop production trend has experienced low yield and has not kept in pace with the annual population growth rate in some countries (Kambewa, 1997; Xavery *et al.*, 2007). Currently Kenya consumes approximately 450,000 tons of beans against a local production level of between 150,000 and 200,000 tons produced annually and imports the deficit mainly from Uganda, Tanzania and Central Africa (MOA, 2011).

Though factors such as low N and acidic soils are important constraints in some parts of Kenya, in the semi-arid areas the soils are less acidic and beans improve soil fertility through biological N-fixation (Giller, 2001) leaving P as the most important constraint for bean production in the dry land areas (Graham *et al.*, 2003; Fabricio *et al.*, 2013). The low soil P and water stress affect bean grain yields and tissue concentration of both Fe and Zn (Astudillo *et al.*, 2008). Studies have shown that the concentration of Fe and Zn in grains and leaves differ (George and Susan, 2010). Decrease in grain and leaf concentration of Fe and Zn has been associated with P toxicity (Gianquinto *et al.*, 2000) or due to dilution effect as a result of P stimulation of growth (Fan *et al.*, 2008; Murphy *et al.*, 2008).

The current study involved three bean genotypes that were developed for canning, these are Awash 1, Awash Melka and Mexican 142. Awash 1 and Awash Melka is a first generation of improved bean genotypes which are more resistant to diseases while Mexican 142 has dominated production as canning beans for over fifty years (Karanja *et al.*, 2011). The response of the three bean genotypes to P application rate and soil moisture on grain yield and tissue concentration of Fe and Zn is unknown.

1.3 Justification of the Study

The three bean genotypes represents some of the promising lines of beans for dry areas and have not been tested on their response to P application rate and soil moisture which is known to affect grain yield and tissue concentration of Fe and Zn (Astudillo *et al.*,2008). The need for beans with high micronutrients is high as either dry beans, green beans or as canning beans due to changing eating habits, preference for fast cooking off-shelf products and high cost of cooking fuel (Karanja *et al.*, 2011). Beans have been grown in Eastern Africa since early 1950s with little work having been done to develop improved bean varieties that combine tolerance to biotic and abiotic stresses with high micronutrient quality. Consequently, bean variety such as Mexican 142 has dominated production despite its susceptibility to Common Bacterial Blight (Tedele, 2006).

The current study contributed in determining the response in grain yield and tissue concentration of Fe and Zn due to P fertilizer application rate and soil moisture in the three beans. Their responses will contribute to the development of agronomic management practices such as P fertilizer application rate and soil moisture requirement which is known to influence productivity of beans and micronutrient quality (Loggerenberg, 2004). In addition, the study contributes in the development of improved bean genotypes that have high grain yield and high micronutrients such as Fe and Zn hence creating an increased stability of beans production (Katungi *et al.*, 2009).

1.4 Objectives

1.4.1 Main Objective

To contribute in determining the effect of P fertilizer application rate on grain yield and tissue concentration of Zn and Fe in three bean genotypes grown under different moisture regimes.

1.4.2 Specific Objectives

1. To determine the effect of P rate on grain yields in three bean genotypes grown under adequate and stressed soil moisture conditions.
2. To evaluate the effect of P rate on grain and leaf tissue concentration of Zn and Fe in three bean genotypes under adequate and stressed soil moisture conditions.
3. To investigate the effect of soil moisture condition on grain and leaf tissue concentration of Zn and Fe in three bean genotypes.

1.4.3 Hypothesis

- H₀:** Phosphorus rate has no difference on grain yield in three bean genotypes under adequate and stressed soil moisture conditions.
- H₀:** Phosphorus application rates have no difference on grain and leaf tissue concentration of Fe and Zn in the three bean genotypes.
- H₀:** Soil moisture regimes have no effect on the grain and leaf tissue concentration of Zn and Fe in three bean genotypes.

CHAPTER TWO

LITERATURE REVIEW

2.1 General Information

2.1.1 Major Producing areas

Common beans originated in Latin America and have two primary centers of origin in the Mesoamerican and Andean regions that are easily distinguished by molecular means (Blair *et al.*, 2006). Major producing countries for national consumption are Brazil and Mexico while the United States, Canada, Argentina and China are all exporting countries. The crop is also important in a range of developing countries of Central America, South America and of Eastern and Southern Africa (Singh, 1999). Beans are grown both for subsistence agriculture and for regional markets where they play an important role in food security and income generation; much of the world's bean production is on small farms ranging from 1-10 acres in size (Katungi *et al.*, 2009)

2.1.2 Common Bean Distribution in Kenya

Common bean production in Kenya is mainly in highland and the midland, about 75 percent of the annual cultivation occurs in three regions namely Rift Valley, Nyanza and Eastern Province (Katungi *et al.*, 2009). In terms of output, the Rift valley contributes the biggest share, accounting for 33 percent of the national output followed by Nyanza (Katungi *et al.*, 2009). Output from eastern parts of the country and the coast is constrained by adverse climatic conditions (Katungi *et al.*, 2009). An impressive high diversity of common bean exists in Kenya having about 80 different cultivars distinguished in different places of the country in late 1970s (Njunguna *et al.*, 1980). The varieties have been losing area because of increased problem of soil infertility such as P deficiency and associated diseases (Ronno *et al.*, 2001).

2.1.3 Bean Farming Requirements

Common bean is a warm season crop that does not tolerate frost or long periods of exposure to near-freezing temperatures at any stage of growth, high temperatures do not affect it if adequate soil moisture is present, although high nocturnal temperatures will inhibit pollination (Katungi *et al.*, 2009). The crop requires moderate amounts of rainfall (300 – 600 mm) but adequate amounts are essential during and immediately after the flowering stage (Gomez, 2004). Generally, common bean is considered a short-season crop with most varieties maturing in a range of 65 to 110 days from emergence to physiological maturity (Buruchara, 2007). Maturity period can continue up to 200 days after planting amongst climbers that are used in cooler upland elevations (Graham and Ranalli, 1997; Gomez, 2004).

2.2 The Role of Fertilizer in Bean Production

Half of the huge yield gaps existing between sub-Saharan African countries and the developed world must be closed through improved soil nutrient management and accompanying field practices while the remainder resolved through widespread adoption of improved crop varieties (Huang *et al.*, 2009). African farmers need better technology that includes more sustainable practices, improved seeds and fertilizer to increase and sustain crops productivity and prevent further degradation of the agricultural lands (Don, 2007). Over exploitation of the soil nutrient has led to P and N deficient soils that are unable to produce a good yield, hence use of fertilizers have become mandatory in farms (Odum, 1989). In order to increase food production in a sustainable manner, farmers will need to use the right fertilizer at the right rate, right time and right place (Norman, 2008).

2.2.1 Role of P on Bean Growth

Beans need P for growth, utilization of sugar and starch, photosynthesis, nucleus formation and cell division, fat and albumen formation, transfer and storage of energy within plants.

Energy from photosynthesis and the metabolism of carbohydrates is stored in phosphate compounds for later use in growth and reproduction (McKenzie and Middleton, 1997; Georgina *et al.*, 2007; Sixbert and George, 2012). Adequate P results in rapid growth, earlier maturity and increased root growth which means plant can explore soil for nutrients and moisture and its deficiency slow overall plant growth, (McKenzie and Middleton, 1997; Sixbert and George, 2012). Crop plants show genotypic variation in P uptake efficiency (i.e. total P uptake), breeding for P-efficient crop cultivars has been recognized as one approach to the management of P-deficient soils (Graham, 1984; Caradus, 1994). Phosphorus and Zn deficiencies are widespread nutritional constraints in bean crop production, and P and Zn interactions have been widely investigated (Marschner, 1995). The genetic improvement of legume P efficiency, deployed in consideration of the social and economic context of local cropping systems, has great potential to address a principal constraint to food security in Africa (Xiaolong *et al.*, 2008).

The current study contributed to the understanding of the responses of the three bean genotypes to varying P application rates and soil moisture regimes on grain yield and tissue concentration of Fe and Zn, which can be utilized in optimizing bean productivity in low soil P farmland (Ann, 2010; Joe *et al.*, 2013).

2.2.2 Root Architecture and P Uptake

Genetic differences exist in the root architecture traits of different bean genotypes that are key adaptations to P stress in low-input agro-ecosystems (Lynch and Brown, 2008). Root traits that enhance topsoil foraging are advantageous in low P soil since P bio-availability is typically greatest in surface horizons (Lynch and Brown, 2001). Genotypes with shallow root architecture have greater growth and yield in low-P soil than related genotypes with deep architectures (Rubio *et al.*, 2003; Zhu *et al.*, 2005). Adventitious roots may improve crop

adaptation to low-P soils by enhancing topsoil foraging (Zhu *et al.*, 2005). In a tropical field study, P stress stimulated adventitious rooting in P-efficient genotypes of common bean (*P. vulgaris*) but not in P-inefficient genotypes. Choosing adventitious rooting is a useful adaptation to low P availability, because adventitious roots explore topsoil horizons more efficiently than other root types (Miller *et al.*, 2003)

Root architectural plasticity traits of *P. vulgaris* that increase topsoil foraging are advantageous for P acquisition but may incur tradeoffs for the acquisition of deep soil resources such as moisture (Lynch and Brown, 2001). In a combined moisture and P stress the genotype that have a dimorphic root system that permit vigorous rooting throughout the soil profile are more advantageous for multiple resource acquisition particularly when resources are differentially localised in the soils (Ho *et al.*, 2005). The roots of plant genotypes that are efficient in mobilizing nutrients from surrounding soil are better able to penetrate and make use of the moisture and minerals contained in subsoil (Susan and George, 2010). These qualities are also associated with greater seedling vigor resulting in increased crop yields (Rengel and Graham, 1995).

2.2.3 The Role of Soil Moisture in P Availability

Inadequate soil moisture lead to poor seed filling resulting into physiological maturity to happen earlier than expected (Brevedan and Egli, 2003; Muasya and Auma, 2003) consequently leading to reduction in grain yield (Szilagyi, 2003). Adequate soil moisture will enhance fertilizer solution and reaction in the soil enhancing plant growth and leading to increased P and other nutrient requirements especially in crops grown under irrigation or in higher rainfall areas (Mckenzie and Middleton, 1997; Georgina *et al.*, 2007). Plants grown in wet soil produced twice as much as those grown on dry soils, nodule weight and activity were five to ten times greater than those from dry soils (Siuet *et al.*, 1984;

Vladimir, 2010).The mineral contents in the seeds are influenced by the soil type and chemical composition and by interaction between genotypes and environment (Moraghan *et al.*, 2002; Cichy *et al.*, 2005). The genotypes by environments interaction has been a large complicating factor in breeding studies with a view toward improvement, principally in bean grain yields, among other traits (Allard and Bradshaw, 1964; Ramalho *et al.*, 1998; Carbonell *et al.*, 2004; Pereira *et al.*, 2009).

2.2.4 Effect of P on Tissue Concentration of Fe and Zn

Studies have shown that increasing the availability of P in the growth medium can induce Zn deficiency in plants by altering soil and plant factors (Robson and Pitman, 1983), but little is known about specific mechanisms. Studies carried out on barley roots observed that Zn deficiency (low tissue Zn concentrations) causes an increase in the expression of P transporter genes in barley roots and also an enhanced P uptake efficiency may cause a decrease in plant uptake of Zn, leading to potentially low Zn concentrations (densities) in food (Huang *et al.*, 2000). A study on grain and straw of winter wheat reported a depressed Zn concentration on increasing P application (Yue *et al.*, 2012). Other studies have shown that an increase in P supply depressed Zn concentration in *P. Vulgaris*, which was attributed to a dilution effect of plant growth (Singh *et al.*, 1988; Gianquinto *et al.*, 2000). Therefore, it remains unclear whether an increase in P availability in the growth medium can reduce Zn uptake by plant roots (Zhu *et al.*, 2001). Many studies have shown that a low Zn supply but a high P supply markedly enhance P concentration in plant tissues, which may cause P toxicity and contribute to symptoms resembling Zn deficiency (Loneragan *et al.*, 1979, 1982; Cakmak and Marschner, 1986; Webb and Loneragan, 1988, 1990).

Study on grain wheat showed that P application significantly decrease grain Zn by 17 to 56 % while the grain levels of Fe, manganese and copper either remained the same or decreased,

also P application increased grain yield but it restricted the accumulation of shoot Zn, but enhanced the accumulation of shoot Fe, Cu and especially manganese (Yue *et al.*, 2012).

2.3 Nutritional Value of Common Beans

Common beans are important for nutritional well-being as well as poverty alleviation among consumers and farmers with few other food or crop options (Broughton *et al.*, 2003). Bean is widely used in the country to make recipe like *Githeri* (cooked mixture of beans and maize) due to increased demand among low income population in the urban areas (Katungi *et al.*, 2009). Common beans provide the crucial proteins (20%), energy (32%) and generous amounts of micro-nutrients especially Fe and Zn, and vitamins A and B complex to over 50 million resource poor rural and urban consumers in eastern Africa (Karanja *et al.*, 2011).

The protein consumption by individual people in the world is estimated at about 77gm of protein per day (FAO, 2010). Recent research also indicates that consumption of grain legumes slows the onset of AIDS in HIV positive people and therefore, an improved bean production would directly address several critical health issues for African communities (Xiaolong *et al.*, 2008). Bean leaves can be used in much the same way that other plant leaves are, as a base for salad or a topping for sandwiches, hamburgers and other foods, they are relatively low in calories and fat, so they may be appropriate choices for dieting individual (Brian, 2011).

2.3.1 Role of Zn and Fe in Human and Plants

Iron is essential for preventing anaemia in human being and for the proper functioning of many metabolic processes while Zn is essential for adequate growth and sexual maturation and for resistance to gastro-enteric and respiratory infections, especially in children (Bouis, 2003). Fe deficiency causes anaemia whose consequences are numerous and grave. Zn deficiency leads to poor child growth, delayed maturation, poor appetite and impaired

immune function(Bouis, 2003). Micronutrient rich common bean cultivars offer unique opportunities for alleviating these disorders in eastern Africa (Karanja *et al.*, 2011).

Plants require the proper balance of Zn for normal growth and optimum yield, Zinc availability in plants depends on soil factors such as the concentration of Zn in solution, ion speciation and the complex interaction of Zn with other macronutrients and micronutrients (Liet *al.*,2003). Selection of cultivars which are rich in Zn can be used in plant breeding to developing high Zn concentration of edible parts without negatively impacting yield(Sadeghzadeh, B. 2013).Zinc deficiency may cause large reductions in crop quality and yield without any visible sign (Alloway, 2004; McDonald *et al.*,2001). Zn deficiency also decreases the amount of Zn in cereal grain and diminishes its nutritional quality (Sadeghzadeh, B.2013). Iron is an essential nutrient for plants whose functions are to accept and donate electrons and also plays important roles in the electron-transport chains of photosynthesis and respiration. Iron is toxic when it accumulates to high levels and act catalytically via the Fenton reaction to generate hydroxyl radicals which can damage lipids, proteins and DNA, hence plants respond to Fe stress in terms of both Fe deficiency and Fe overload (Erin and Connolly, 2002; Goff *et al.*, 2002).

2.3.2 Source and Sink Strength in Plant Growth

Studies have shown that the sink strength which depend of sink age rather than sink size may determine the partitioning of plants assimilates in the plant organ(Marcelis, 1996). Information on the effect of source strength on the partitioning of assimilates among the plant organs is limited (Ho, 1988, 1992). However, studies show hierarchy among sink in different cultivars that is some (e.g. fruits, seeds or underground storage organs) have priority and suffer less from a reduction in assimilate supply than other organs e.g. flowers. In relation to

in assimilate supply, moisture condition affects the net photosynthesis, hence the growth and the supply of assimilates and photosynthates (Wardlaw, 1990).

CHAPTER THREE

MATERIALS AND METHODS

3.1 Experimental Site

The experiment was carried out between August and November 2012 at KARI, Katumani in Machakos located at 1°35'S: 37°14'E, and 1560 masl. The Centre experiences a semi-arid tropical climate described as AEZ 3 (Jaetzold and Schimidt, 1983), with a bimodal pattern of rainfall. The long rains are received between March and June, with the peak in April, followed by a dry period that extends to mid-October. The short rains begin in mid-October, peak in November and taper off towards mid-December.

3.2 Bean Genotypes

Three genotypes used in the study were Awash 1, Awash Melka and Mexican 142. Awash 1 and Awash Melka is a first generation of improved bean genotypes, a promising line of micronutrient dense and disease resistant beans for canning that originated from Ethiopia. Mexican 142 has low grain yield and susceptible to diseases such as Common Bacterial Blight and has dominated production as canning beans for over 50 years (Karanja *et al.*, 2011). The three bean genotypes were assigned letters as follows Awash 1 (V_1), Awash Melka (V_2) and Mexican 142 (V_3).

3.3 Experimental Design Layout

The experiment was laid out in a RCBD with treatments in a split-split plot arrangement with three replicates, comprising moisture regimes as main plots, the three bean genotypes as the sub plots and five P fertilizer application rates as sub-sub plots. The five rates of P fertilizer application were 0 kg ha⁻¹, 20 kg ha⁻¹, 40 kg ha⁻¹, 60 kg ha⁻¹ and 80 kg ha⁻¹. Each sub plot measured 1.5 m x 1.5 m. Soil analysis was done before the start of the experiment and

indicated that it was sandy loam of Ferric Luvisol origin as described by Kibe *et al.*, (1981), it was also slightly acidic with low total N, organic carbon and P.

Two bean seeds were sown per hill with 20 cm spacing within rows and 50 cm between rows and thinned to one plant per hill two weeks after germination. Main plot was separated by a one meter foot path for easy access during data collection and harvesting. The plots were kept weed free by hand weeding. To prevent cutworms, Sulban[®]48ec (Chlorpyrisos 480g/l) insecticide was applied after germination. Dimethoate[®]40ec (Dimethoate 400g/l) insecticide was applied to control white flies on emergence. Soil samples were randomly obtained prior to sowing from the top soil at increments of 10cm upto a depth of 30cm for chemical analysis as described (Hinga *et al.*, 1980).

The amount of moisture that the soil can hold at field capacity was determined using gravimetric method prior to starting the experiment. After flowering, moisture stress was applied to one of the main plot selected randomly at 70% of field capacity by watering once in two weeks. Watering was done using drip irrigation by laying drip lines along each bean rows (Each sub-subplot served by four drip lines). A 5000 liter water tank was placed near the farm to act as water reservoir throughout the growing season. A main pipe connected to the water tank and fitted with valves to regulate flow of water supplied water to the drip lines. To prevent blockage of the drip lines, care was taken to prevent dust and soil particles from falling on the drip holes during laying and subsequent weeding.

3.4 Data Collection

3.4.1 Temperatures and Days to 50% Flowering

Minimum and maximum temperatures were recorded from the meteorological station using a wet and dry bulb thermometer. The number of days to 50% flowering was recorded for each P application rate to determine the period for taking the leaf sample for Fe and Zn test.

3.4.2 Leaf Sampling for Zn and Fe Analysis

At 50% flowering, three fully expanded leaves were picked from six plants grown in the two middle rows (except those grown on the end of each) for Fe and Zn test samples making a total of 12 plants sampled per each sub-sub plot (each sub-plot had a total of 32 plants). The leaves picked were initially washed in distilled water, then hydrochloric acid and distilled water to remove any impurities that may have longed on the leaves and finally rinsed in distilled water and sun dried for two hours to remove dripping water which can damage Khaki paper envelope where it was later stored. The sample was then oven dried for 48 hours at a temperature of 60°C and stored in khaki paper for analysis.

3.4.3 Harvesting at Bean Maturity

At harvest time, pods were picked from each plant in the two middle rows, in each sub-sub plot, leaving one plant at the two ends of each row to avoid the border effects. The harvested beans were threshed and the grains weight taken, this was followed by drying the beans at 60°C for 48 hours. The weight of the dried beans was recorded for each sample. In order to obtain a 5 gram representative sample for micronutrient analysis, the dried grains from each plot were evenly placed on clean acid (Hydrochloric acid 0.1%) washed tray and spread in a circle of 15cm diameter then divided into four quarters of equal size. A sample of 5 grams was collected randomly from each quarter making a total of 20 grams. This was followed by a quartering procedure repeated until a total of 5 gram representative sample was obtained.

The 5 gram samples obtained for both leaf and grain sample were grounded using a non-contaminating grinding Retsch mill with Teflon chambers and Zirconium balls to avoid Fe and Zn contamination. A sample of 0.5–0.8 grams was collected randomly for analysis from each of the 5gram sample through a quartering procedure. The grinded samples were packed and labeled in clean plastic screw-top tubes and stored in a clean, dry and insect-free location. Analysis was done using Inductively Coupled Plasma–Optical Emission Spectroscopic (ICP-OES).

3.5 Data Analysis

The data on mean grain weight, grain and leaf tissue concentration Fe and Zn were analysed using ANOVA(SAS 8.2) to isolate treatment effect of P and moisture. The mean values were separated using LSD at $P < 0.05$ significant level. Correlation analysis between grain and leaf tissue concentration of Fe and Zn, as well as grain weight to leaf and grain tissue concentration of Fe and Zn were determined using Pearson Correlation (SAS 8.2).

CHAPTER FOUR

RESULTS

4.1.1. Effect of P rate on Grain Yield of Three Bean Genotypes under Adequate and Stressed Moisture Conditions

Bean grain yields showed a significant ($df = 16; F = 3.19; P = 0.0006$) interaction between P application rates, moisture regimes and genotypes (Table 2). Under both adequate and stressed moisture conditions, bean grain yields of the three bean genotypes increased significantly ($df = 28; t = 2.05; P < 0.05$) with increased P fertilizer application rate up to 60 kg ha⁻¹, above which there was no increase (Table 1). Genotype V₂ produced significantly ($df = 28, t = 2.05; P < 0.05$) higher grain yields than all the others under adequate moisture but under moisture stress it was similar to V₃ but higher than V₁. The three bean genotypes produced significantly ($df = 58; t = 2; P < 0.05$) higher grain yield under adequate soil moisture than under stressed soil moisture.

Table 1: Grain Yields (Kg Ha⁻¹) of Three Bean Genotypes (V₁, V₂, V₃) Under Varying P Rate in Adequate and Stressed Moisture Conditions

Moisture regimes	Genotypes	P rate (kg ha ⁻¹)					Mean
		0	20	40	60	80	
Adequate	V ₁	2242.33Dc	2868.11Cb	3536.53Ba	4036.67Aa	4065.13Aa	3349.8b
	V ₂	2774.70Da	3155.40Ca	3574.46Ba	3873.13Ab	3953.70Ab	3466.4a
	V ₃	2550.46Db	2743.40Cc	2932.10Bb	3318.47Ac	3306.60Ac	2970.4c
	Mean	2522.7D	2922C	3348B	3743A	3775.3A	3262.2¹
	LSD	Rows = 180.25					
	SE	Rows ±62.22					
Stressed	V ₁	1171.86De	1477.67Ce	1602.83Bd	1787.30Ae	1825.60Ae	1573.0e
	V ₂	1320.77Dd	1531.23Cd	1764.50Bc	1883.46Ad	1931.83Ad	1686.60d
	V ₃	1254.37Dd	1576.27Cd	1735.10Bc	1877.30Ad	1952.70Ad	1679.00d
	Mean	1249.00Dd	1528.30C	1701.00B	1849.30A	1903.70A	1646.20²
	LSD	Rows = 122.42					68.20
	SE	Rows ±42.2586					
Grand Mean		1885.80D	2225.2C	2524.50B	2796.20A	2839.50A	
LSD		Row 107.81, Across all columns 83.51					
SE		last row±38.09; Across all columns ±29.5					

Grain yield in the same row followed by the same upper case letters(A,B...D) or in the same column followed by the same lower case letters(a, b, c...e)or superscript numerical letter(1,2) are not significantly ($P < 0.05$) different.

4.1.2The Effect of P Rate on Grain Fe(Mg Kg⁻¹) of Three Bean Genotypes under Adequate and Stressed Moisture Conditions

Bean grain Fe concentration showed a significant ($df = 16$; $F = 5.47$; $P < 0.0001$) interaction between P application rates, moisture regimes and genotypes (Table 2).

Grain Fe concentration significantly ($df = 28$; $t = 2.05$; $P < 0.05$) increased with P application rate up to 40kg ha^{-1} in all the three bean genotypes irrespective of the soil moisture condition. An increase in P fertilizer rate above 40kg ha^{-1} led to a decrease in grain Fe in all the three genotypes under moisture adequate condition. Under the moisture stressed condition a decrease in grain Fe in V_1 and V_2 at rate above 40kg ha^{-1} was noted while V_3 showed a decrease at P application above 60kg ha^{-1} (Table 2). Genotype V_2 had highest grain Fe concentration irrespective of moisture conditions followed by V_3 whilst genotype V_1 showed the lowest grain Fe concentration.

Table 2: Grain Fe (Mg Kg⁻¹) Concentration of Three Bean Genotypes in Varying P Application Rate under Adequate and Stressed Moisture Condition

Moisture regimes	Genotypes	P rate (kg ha ⁻¹)					Mean
		0	20	40	60	80	
Adequate	V ₁	61.2Ed	68.8Be	71.7Ae	66.0Ce	63.0De	66.0e
	V ₂	80.7Ea	84.8Da	92.4Aa	90.0Ba	83.1Ca	86.0a
	V ₃	71.4Dc	75.0Cc	82.6Ac	81.4Bc	72.1Dd	77.0c
	Mean	71.0E	76.0C	82.0A	79.0B	72.0D	76.3 ¹
	LSD	Rows = 1.5408					
	SE	Rows ±0.5319					
Stressed	V ₁	57.1Ce	63.8Bf	66.2Af	64.5Bf	58.3Cf	62f
	V ₂	77.6Db	78.8Db	87.1Ab	82.6Bb	80.7Cb	81b
	V ₃	70.4Ec	73.8Dd	78.6Ad	78.8Ad	76.7Cc	76d
	Mean	68E	72C	77A	75B	72C	73 ²
	LSD	Rows = 1.2251					0.6063
	SE	Rows ±0.4229					
Grand Mean		69.7E	74.2C	79.8A	77.2B	72.3D	74.64
LSD		Row 0.9586, Across all columns 0.7425					
SE		last row ±0.3386; Across all columns ±0.2623					

Grain Fe concentration in the same row followed by the same upper case letters (A, B...D) or in the same column followed by the same lower case letters (a, b, c...e) or superscript numerical letter (1,2) are not significantly ($P < 0.05$) different.

4.1.3 The Effect of P Application rates on Bean Grain Zn (Mg Kg⁻¹) Concentration in Three Bean Genotypes under Adequate and Stressed Moisture Conditions

The grain Zn concentration had a significant ($df = 16$; $F = 6.67$; $P < 0.0001$) interaction between soil moisture, P application rate and genotypes. Under moisture adequate condition,

genotype V₁ and V₂ showed significant ($df = 28; t = 2.05; P < 0.05$) increase in grain Zn concentration up to 40 kg ha⁻¹ P (Table 3), but P fertilizer application rate above 40 kg ha⁻¹ led to a decrease in grain Zn concentration in genotypes V₂ and V₃.

Similarly, under moisture stressed condition, P fertilizer application significantly ($df = 28; P < 0.05$) increased grain Zn concentration up to 40 kg ha⁻¹ in genotype V₁, but in genotype V₂ it increased up to 60 kg ha⁻¹. Genotypes V₂ and V₃ had the highest grain Zn concentration compared to V₁ under moisture stressed condition (Table 4). The grain Zn concentration showed no significant ($df = 58; t = 2.0; P < 0.05$) difference.

Table 3: Grain Zn (Mg Kg⁻¹) Concentration of Three Bean Genotypes in Varying P Application rate under Adequate and Stressed Moisture Condition

		P rate (kg ha⁻¹)					
Moisture regimes	Genotypes	0	20	40	60	80	Mean
Adequate	V ₁	18.9Cd	19.6Be	19.8Ae	20.3Ac	19.6Bd	19.5b
	V ₂	22.2Db	24.3Bc	25.5Aa	24.4Ba	23Cb	23.9a
	V ₃	20.2Dc	24.7Ab	25.2Ab	24.3Ba	23.9Ca	23.7a
	Mean	20.4C	22.9A	23.5A	23A	22.2B	22.4¹
	LSD	_____ Rows = 0.616 _____					
	SE	_____ Rows ±0.2126 _____					
Stressed	V ₁	19.1Cd	19.6BCe	20.9Ad	19.9Bd	19.3BCd	19.8b
	V ₂	23Da	23.8Cd	24.2BCc	24.4Aa	23Db	23.7a
	V ₃	22.1Cb	25.7Aa	25.9Aa	23.4Bb	21.5Dc	23.7a
	Mean	21.4D	23B	23.7A	22.6B	21.3C	22.4¹
	LSD	_____ Rows = 0.5545 _____					<u>0.2578</u>
	SE	_____ Rows ±0.1914 _____					
Grand Mean		20.9D	23B	23.6A	22.8B	21.7C	22.4
LSD		_____ Row 0.4077, Across all columns 0.3158 _____					
SE		_____ Last row ±0.1440; Across all columns ±0.1115 _____					

Grain Zn concentration in the same row followed by the same upper case letters (A, B...D) or in the same column followed by the same lower case letters (a, b, c...e) or superscript numerical letter (1, 2) are not significantly ($P < 0.05$) different.

4.2.1 The Effect of P Application rate on Leaf Fe Concentration of Three Bean Genotypes under Adequate and Stressed Moisture Conditions

Bean leaf Fe concentration showed highly significant ($df = 16$; $F = 82.76$; $P < 0.0001$) interaction between P fertilizer application rate, genotypes and moisture regimes. Under adequate moisture conditions, leaf Fe concentration in genotype V₂ and V₃ significantly ($df = 28$; $t = 2.05$; $P < 0.05$) decreased with increasing rate of P application up to 60 kg ha⁻¹ above which there was no significant ($df = 28$; $t = 2.05$; $P < 0.05$) decrease in leaf Fe concentration in all the three bean genotypes (Table 5). Under adequate moisture conditions, genotype V₂ had highest (overall) leaf Fe concentration while genotype V₃ had the highest leaf Fe concentration under stressed moisture condition. Under stressed moisture condition, leaf Fe concentrations significantly ($df = 28$; $t = 2.05$; $P < 0.05$) decreased with increasing rate of P application up to 40 kg ha⁻¹ of P application (Table 4). There was a significantly ($df = 58$; $t = 2.0$; $P < 0.05$) higher leaf Fe concentration in bean grown under stressed water condition than in those grown under adequate water condition.

Table 4: Leaf Fe Concentration of Three Bean Genotypes in Varying P Application Rate under Adequate and Stressed Moisture Conditions

		P rate (kg ha⁻¹)					
Moisture regimes	Genotypes	0	20	40	60	80	Mean
Adequate	V₁	291Ae	278Bd	239Dd	242Cd	241Cd	258e
	V₂	316Ad	285Bc	267Cb	262Db	261Db	278b
	V₃	332Ac	305Bb	244Cc	233De	232De	269d
	Mean	313A	289B	250C	246D	245D	268²
	LSD	_____ Rows = 1.5384 _____					
	SE	_____ Rows ±0.5310 _____					
Stressed	V₁	341Ab	288Bc	243Ec	258Cc	250Dc	276c
	V₂	256Af	242Be	227Ce	230Cf	230Ce	237f
	V₃	370Aa	352Ba	294Da	302Ca	285Ea	321a
	Mean	322A	294B	255C	263D	255C	278¹
	LSD	_____ Rows = 3.7741 _____					_1.2431_
	SE	_____ Rows ± 1.3028 _____					
Grand Mean		318.3A	292B	252.7C	255.7D	250.6E	273.9
LSD		_____ Row 1.9655, Across all columns 1.5224 _____					
SE		_____ Last rows ±0.6943; Across all columns ±0.5378 _____					

Leaf Feconcentration in the same row followed by the same upper case letters(A,B...D) or in the same column followed by the same lower case letters (a, b, c...e)or superscript numerical letter(1,2)are not significantly different ($P<0.05$).

4.2.2 The Effect of P Application rate on Leaf Zn Concentration of Three Bean Genotypes under Adequate and Stressed Moisture Conditions

Leaf Zn concentration showed a significant ($df = 16$; $F = 2.95$; $P = 0.0013$) interaction between P rate, moisture conditions and genotypes. Under adequate moisture condition, genotype V₁ and V₂ had a significant ($df = 28$; $t = 2.05$; $P < 0.05$) decrease in leaf Zn at P application up to 40 kg ha⁻¹ and there was no significant ($df = 28$; $t = 2.05$; $P < 0.05$) difference in P application above 60 kg ha⁻¹ in the three bean genotypes (Table 5).

Under moisture stressed condition, leaf Zn concentration significantly ($df = 28$; $t = 2.05$; $P < 0.05$) decreased up to 40 kg ha⁻¹ of P application in genotype V₁ and V₂. Genotype V₃ had the highest leaf Zn concentration under moisture stressed condition (Table 5). The three bean genotypes had significantly ($df = 58$; $t = 2.0$; $P < 0.05$) higher leaf Zn concentration when grown under stressed moisture condition than under adequate moisture condition.

Table 5: Leaf Zn Concentration of Three Bean Genotypes in Varying P Application rate under Adequate and Stressed Moisture Conditions

		P rate (kg ha⁻¹)						
Moisture regimes	Genotypes	0	20	40	60	80	Mean	
Adequate	V ₁	31.5Ab	29.2Bb	27.2Cc	28.3Bb	28.0Ba	29.1b	
	V ₂	30.4Ac	28.3Bc	26.5Cd	23.5Cd	23.3Cd	26.4e	
	V ₃	29.7Ad	29.3Ab	26.7Bc	26Bc	26.3Bb	27.6d	
	Mean	30.5A	28.9B	26.8C	25.9D	25.9D	27.7²	
	LSD	_____Rows = 0.9316_____						
	Std Error	_____Rows ±0.3216_____						
Stressed	V ₁	30Acd	29Bb	28.9Cb	28.3Cb	26.2Dc	28.5c	
	V ₂	30.5Ac	29.7Ba	28.7Cb	28.3Cb	28Ca	29bc	
	V ₃	33.3Aa	30Ba	29.6Ba	29.2Ca	26.8Db	29.8a	
	Mean	31.3A	29.6B	29.1BC	28.6C	27D	29.1 ¹	
	LSD	_____Rows =0.9474_____						__0.4159__
	SE	_____Rows ± 0.3270_____						
Grand Mean		30.9A	29.25B	27.9C	27.3C	26.4D	28.4	
LSD		_____Row 0.6575, Across all columns 0.5093_____						
SE		_____Last row 0.2323; Across all columns ±0.1799_____						

Leaf Zn concentration in the same row followed by the same upper case letters(A,B...D) or in the same column followed by the same lower case letters (a, b, c...e) or superscript numerical letter(1,2)are not significantly ($P<0.05$) different.

4.2.3. Effect of P Application rate on Percentage of Leaf Fe Accumulated in Grain under Adequate and Stressed Moisture Condition in Three Bean Genotypes

The results for the percentage of leaf Fe accumulated in grain indicated a highly significant ($df = 16$; $F = 13.36$; $P < 0.0001$) interaction between P fertilizer application rates, genotypes and moisture regimes. Under adequate moisture conditions, an increase in P application up to 40 kg ha^{-1} led to a significant ($df = 28$; $t = 2.05$; $P < 0.05$) increase in percentage leaf Fe accumulated in grain in genotypes V_1 and V_2 whilst in genotypes V_3 , percentage accumulation of leaf Fe in grain increased significantly ($df = 28$; $t = 2.05$; $P < 0.05$) with increasing rate of P application up to 60 kg ha^{-1} and declined (Table 6). Under stressed moisture condition, percentage of leaf Fe accumulated in grain increased significantly ($df = 28$; $t = 2.05$; $P < 0.05$) with increasing P application up to 40 kg ha^{-1} in all genotypes. Genotype V_2 had the highest percentage of leaf Fe accumulated in grain under stressed moisture condition (Table 6). Beans grown under moisture adequate condition had significantly ($df = 58$; $t = 2.0$; $P < 0.05$) higher percentage of leaf Fe accumulated in grain Fe than those grown under moisture stressed conditions (Table 7).

Table 6: Percentage (%) of Leaf Fe Accumulated to Grain of Three Bean Genotypes in Varying P Application Rate under Adequate and Stressed Moisture Conditions

		P rate (kg ha⁻¹)						
Moisture regimes	Genotypes	0	20	40	60	80	Mean	
Adequate	V ₁	21Dc	25Cc	30Ad	26Bd	26Bd	26d	
	V ₂	26Eb	30Db	35Ab	34Bc	32Ca	31b	
	V ₃	21Ec	25Dc	34Bc	35Ab	32Ca	29c	
	Mean	23E	27D	33A	32B	30C	29 ¹	
	LSD	_____ Rows = 0.9265 _____						
	SE	_____ Rows ±0.3198 _____						
Stressed	V ₁	17E	22Dd	27Ae	25Be	23Ce	23f	
	V ₂	31Da	33Ca	38Aa	36Ba	31Db	35a	
	V ₃	19D	21Ce	27Ae	26Bd	27Ac	24e	
	Mean	22D	25C	31A	29B	28.5B	27 ²	
	LSD	_____ Rows = = 0.5727 _____						_.04159_
	SE	_____ Rows ±0.1977 _____						
Grand Mean		23E	26D	32A	30B	29C	28	
LSD		_____ Row 0.5267, Across all columns 0.408 _____						
SE		_____ Last row ± 0.1861; Across all columns ±0.1441 _____						

Percentage (%) of leaf Fe accumulated to grain in the same row followed by the same upper case letters(A,B...D) or in the same column followed by the same lower case letters (a, b, c...e) or superscript numerical letter(1,2)are not significantly ($P < 0.05$) different.

4.2.4 Effect of P Application Rate on Percentage of Leaf Zn Accumulated to Grain under Adequate and Stressed Moisture Condition in Three Bean Genotypes

A highly significant ($df = 16$; $F = 4.16$; $P < 0.0001$) interaction between P application rates, genotypes and moisture regimes on percentage of Leaf Zn accumulated to grain was observed. Under adequate moisture condition, percentage of leaf Zn accumulated to grain increased significantly ($df = 28$; $t = 2.05$; $P < 0.05$) with increasing rate of P application up to 40 kg ha⁻¹ except genotype V₂ which increased up to 60 kg ha⁻¹ (Table 7).

Under moisture stress condition, percentage of leaf Zn accumulated to grain in genotype V₁ and V₂ increased significantly ($df = 28$; $t = 2.05$; $P < 0.05$) with increasing rate of application up to 40 kg ha⁻¹. Under moisture stress condition, genotype V₂ and V₃ had the highest percentage of leaf Zn accumulation to grain (Table 7). Beans grown under moisture adequate condition had significantly ($df = 58$; $t = 2.0$; $P < 0.05$) higher percentage of leaf Zn accumulated to grain than those grown under moisture stressed conditions (Table 7).

Table 7: Percentage(%) of Leaf Zn Accumulated in Grain of Three Bean Genotypes in Varying P Application rate under Adequate and Stressed Moisture Conditions

		P rate (kg ha⁻¹)					
Moisture regimes	Genotypes	0	20	40	60	80	Mean
Adequate	V ₁	60Cf	67.3Bc	72.3Ae	71.6Ae	71Af	68.4d
	V ₂	73.2Db	85.7Ca	96.2Ba	100Aa	98Aa	90.6a
	V ₃	68.2Dc	84.7Ca	94.2Ab	93.6Ab	90.8Bb	86.3b
	Mean	67.1C	79.2B	87.6A	88.4A	86.6A	81.8 ¹
	LSD	_____Rows = 2.856_____					
	SE	_____Rows ±0.9859_____					
Stressed	V ₁	63.7Ce	67.7Bc	72.3Ae	70.5Be	73.9Ae	69.6d
	V ₂	75.4Ca	80.1Bb	84.3Ad	86.5Ac	82.3Bc	81.7c
	V ₃	66.4Cd	85.7Aa	87.6Ac	80.2Bd	80.3Bd	80c
	Mean	68.5C	77.8B	81.4A	79.1A	78.8A	77.1 ²
	LSD	_____Rows = 3.2141_____					_____1.3278_____
	SE	_____Rows ± 1.1095_____					
Grand Mean		67.8C	78.5B	84.5A	83.7A	82.4A	79.4
LSD		_____Row 2.0994, Across all columns 1.6262_____					
SE		_____Last row ±0.7416; Across all columns ±0.5745_____					

Percentage (%) of leaf Zn accumulated to grain in the same row followed by the same upper case letters(A,B...D) or in the same column followed by the same lower case letters (a, b, c...e) or superscript numerical letter(1,2)are not significantly ($P<0.05$) different.

4.2.5. Comparison of Leaf and Grain Concentration of Fe in Three Bean Genotypes under Adequate and Stressed Moisture Conditions

All genotypes had higher mean leaf Fe concentration than grain Fe concentration under adequate moisture condition. Leaf Fe concentration followed the order $V_2 > V_3 > V_1$, while the grain Fe concentration followed $V_2 > V_3 > V_1$ (Fig.1) All genotypes had higher mean leaf Fe concentration than grain Fe under stressed moisture condition. Leaf Fe concentration followed the order $V_3 > V_1 > V_2$, while the grain Fe concentration followed $V_2 > V_3 > V_1$ (Fig. 2)

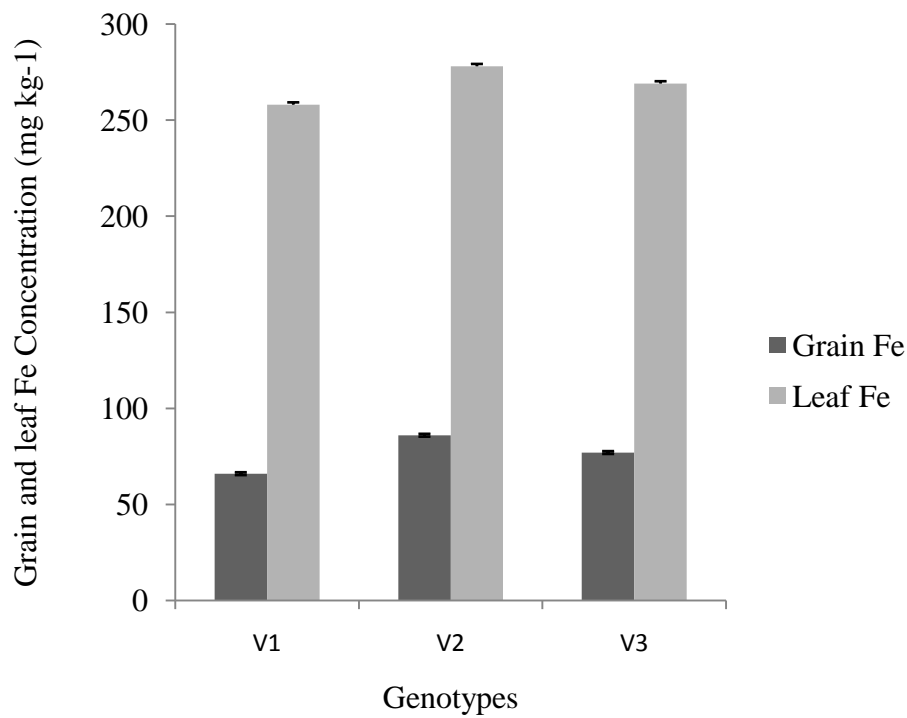


Figure 1: Comparison of leaf and Grain Concentration of Fe in Three Bean Genotypes under Adequate Moisture Condition.

NB: Error bar represent LSD ($P < 0.05$) for the comparison of genotypes at each P rates.

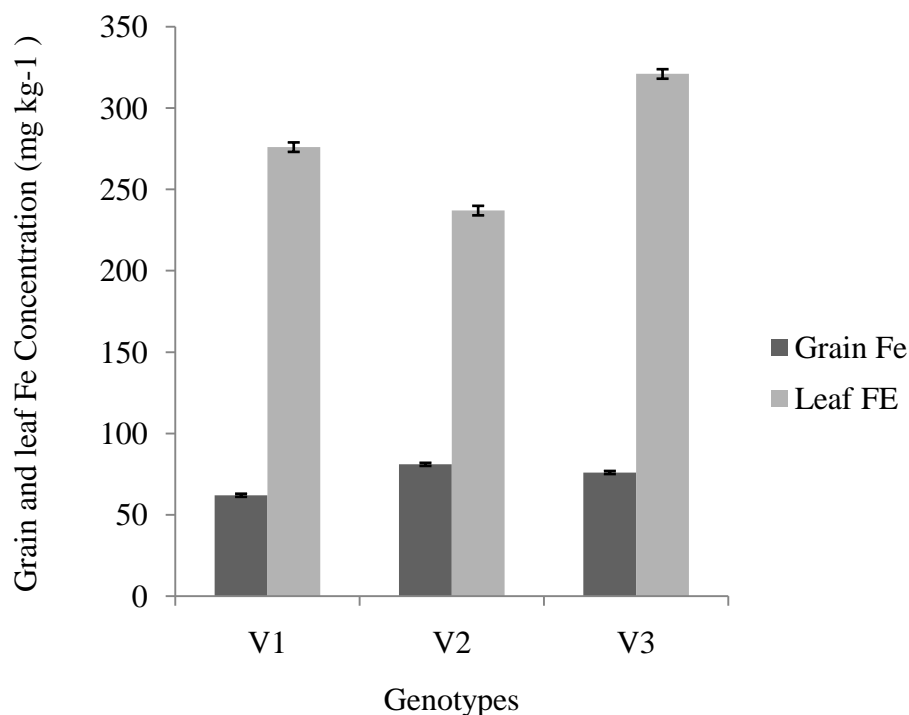


Figure 2: Comparison of Leaf and Grain Concentration of Fe in Three Bean Genotypes under Stressed Moisture Condition

NB: Error bar represent LSD ($P < 0.05$) for the comparison of genotypes at each P rates.

4.2.6. Comparison of Leaf and Grain Concentration of Zn in Three Bean Genotypes under Adequate and Stressed Moisture Conditions

Zinc concentration was higher in leaf than in grain in all genotypes. Leaf Zn concentration followed the order $V_1 > V_3 > V_2$, while the grain Zn concentration followed the order $V_2 = V_3 > V_1$ (Fig.3) under adequate moisture condition. Zinc concentration was higher in leaf than in grain in all genotypes. Leaf Zn concentration followed the order $V_3 = V_2, V_2 = V_1$ while the grain Zn concentration followed the order $V_2 = V_3 > V_1$ under stressed moisture conditions (fig 4).

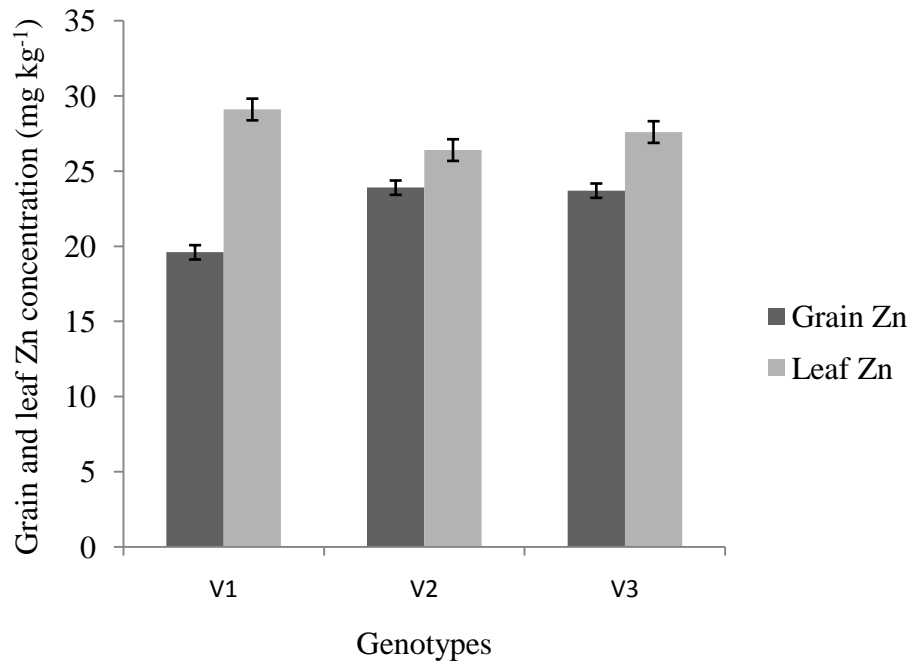


Figure3: Comparison of Leaf and Grain Concentration of Zn in Three Bean Genotypes under Adequate Moisture Condition.

NB: Error bar represent LSD ($P < 0.05$) for the comparison of genotypes at each P rates.

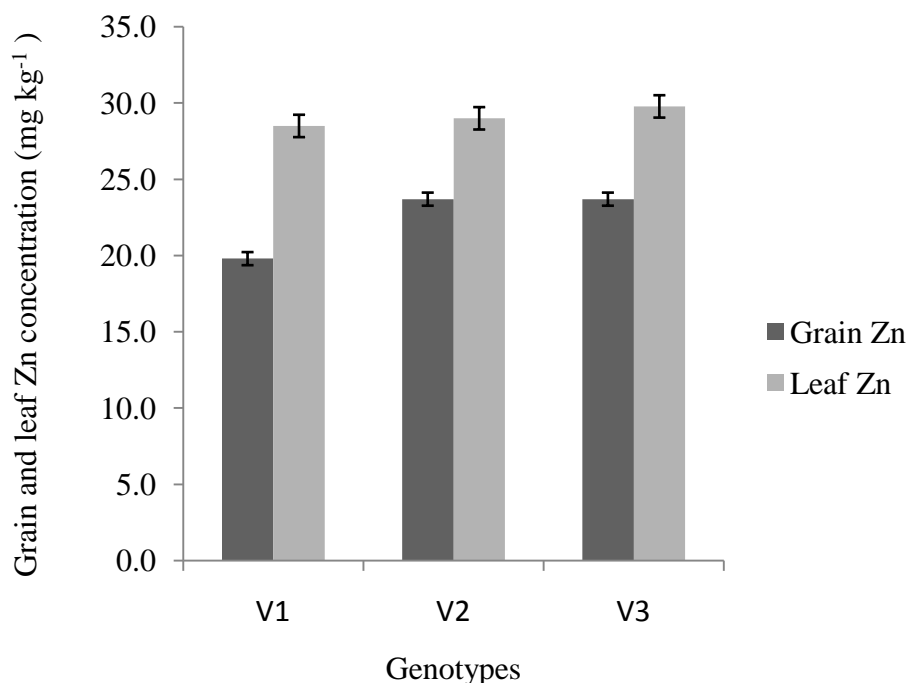


Figure 4: Comparison of Leaf and Grain Concentration of Zn in Three Bean Genotypes under Stressed Moisture Condition

NB: Error bar represent LSD ($P < 0.05$) for the comparison of genotypes at each P rates.

4.2.7. Correlation between Grain Yield and Leaf and Grain Tissue Concentration of Fe and Zn

A highly significant ($n = 90; r = 0.79736; P < 0.0001$) positive correlation between the grain Fe and Zn concentration and a highly significant ($n = 90; r = 0.53662; P < 0.0001$) positive correlation between leaf Fe and Zn concentration were observed. A highly significant ($n = 90; r = -0.34860; P = 0.0008$) negative correlation between the grain yield and leaf Fe concentration and highly significant ($n = 90; r = -0.58292; P = 0.0001$) negative correlation between grain yield and leaf Zn concentration were observed (Table 8)

Table 8: Correlation between Grain Yield, Leaf and Grain Concentration (Mg kg⁻¹) of Fe and Zn

Concentration	Grain yield	grain Fe	grain Zn	leaf Fe
Grain Fe	0.24686*			
Grain Zn	0.06638ns	0.79736**		
Leaf Fe	-0.34860*	-0.20375ns	-0.06295ns	
Leaf Zn	-0.58292**	-0.32753*	-0.16428ns	0.53662**

Significant (P < 0.05); ** highly significant (P < 0.01); ns - not significant (P > 0.05).
 .|r| Pearson Correlation Coefficients

CHAPTER 5

DISCUSSION, CONCLUSION AND RECOMMENDATIONS

5.1 DISCUSSION

5.1.1 Effect of P Application on Grain Yields in Three Bean Genotypes under Adequate and Stressed Moisture Conditions

The current study showed that increase in P application rate led to an increase in grain yield up to the optimal level of 60kg ha⁻¹ under adequate and stressed soil moisture conditions in all the bean genotypes. This shows that grain yield increase in the three beans is a combined effect of P application, soil moisture regime and genotypes. The significant grain yield increase with increasing P fertilizer in beans grown under adequate moisture regime in the current study implies that so long as soil conditions are favorable, grain yield will significantly increase with increasing P application up to an optimal level. Other studies indicated a significant increase in grain yield with increasing P application up to optimal levels (Al- Farha and Al-Rawi, 2002; Kamanga *et al.*, 2010; Davood, 2013).

Adequate P results in rapid growth, earlier maturity and increased root growth which means plant can explore soil for nutrients and moisture and its deficiency slow overall plant growth, (McKenzie and Middleton, 1997; Georgina *et al.*, 2007). This process of nutrient uptake by plants has been associated with increased root growth enabling plants to explore more soil nutrients leading to an increased grain yield (Ho *et al.*, 2005; Fageria and Stone, 2006; Fageria *et al.*, 2006). The increase in the grain yield with increasing rate of P application in the current study may be attributed to increased root growth due to increasing P application rate leading to increase nutrient uptake hence increased grain yield.

In the current study bean grown in soil moisture adequate conditions produced more grain yield than those grown in soil moisture stressed conditions irrespective of the P application rate and genotypes. This corroborates other studies that showed that regardless of the P rate of application and superiority of the bean genotype, inadequate soil moisture led to poor seed filling resulting into physiological maturity to happen earlier than expected (Brevedan and Egli, 2003; Muasya and Auma, 2003) eventually leading to reduction in grain yield (Szilagyi, 2003). Crops have marked moisture sensitive stages of which if moisture stress occurs during this stage, crop growth and development is hindered leading to reduced yield, hence bean crop growth needs to be synchronized to soil moisture and P rate to get maximum seed yield (Shah *et al.*, 2001; Cassman and Waters, 2002).

The current study shows genotype variation in the grain yield responses regardless P rate of application and moisture regime. The genotype variations imply that choosing superior bean genotype should be considered in order to increase grain yield irrespective of rate of P application and moisture regime. The current study corroborate other finding that emphasized the importance of choosing bean genotypes that have high grain yield and had superiority in P acquisition efficiency (Zhang, 2007; Fageria *et al.*, 2008; Hammond *et al.*, 2008). Overall, the current study emphasizes the importance of not only correction of P deficiency through P fertilizer application rate of 60kg ha⁻¹, ensuring adequate soil moisture is maintained as well as growing high yielding superior genotypes Awash Melka (V₂) in order to have increased bean grain yield.

5.1.2 Effect of P Application and Moisture Stress on Grain Concentration of Zn and Fe in Three Bean Genotypes

Grain Fe and Zn concentration in the three bean genotypes significantly increased due to a combined effect of moisture regime, P rates and genotypes in the current study. The process

of P acquisition in plants has been associated with root architecture development (Miller *et al.*, 2003; Liao *et al.*, 2004; Beebe *et al.*, 2006). A favorable moisture condition improves root development leading to increased scavenging of soil mineral resources such as Fe and Zn in P efficient genotypes (Hoet *et al.*, 2005). An increase in P application in bean genotype under favorable moisture condition led to increase in grain Fe and Zn concentration up to P optimal level, such increase was characterized by genotype variations. The genotype variability observed in the current study corroborates a study finding by Astudillo *et al.*, (2008) that showed genotype variability in beans response to P rate of application on grain and leaf concentration of Fe and Zn.

Under adequate moisture condition, P application above the optimal level led to a decrease in Fe and Zn concentration irrespective of the genotype. This is associated with dilution effect as a result of P stimulation of growth leading to increased grain yield with increased P application (Fan *et al.*, 2008; Murphy *et al.*, 2008). Other studies have attributed such decrease in grain Fe and Zn to toxicity effect (Gianquinto *et al.*, 2000) or due to complex interaction of P with other minerals such as Mn, Fe which is known to occur under different cultural systems (Barben *et al.*, 2011). The optimal P level at which the grain concentration of Fe and Zn begins to depress in the current study may be attributed to the low native soil P level of 15 mg kg⁻¹ at Katumani field unlike soils with high P especially in countries like China with soil P level of 24 mg kg⁻¹ (Li *et al.*, 2011), East Asia and West Europe (MacDonald *et al.*, 2011) which has experienced continuous P fertilizer applications from recent decades, resulting in cumulative surplus of P in croplands. In areas with high soil P levels a depressed grain concentration of Fe and Zn can be attained at a low soil P levels (Yue *et al.*, 2012; Ryan *et al.* 2008; Astudillo *et al.*, 2008). Irrespective of P application, beans grown under adequate moisture condition produced an overall higher grain Fe concentration

than those grown under stressed moisture condition; this is attributed to the role of moisture in uptake of soil nutrients (Priscilao *et al.*, 2008).

5.1.3 Effect of P Fertilizer Application and Moisture regime on Bean Leaf Fe and Zn Concentration

A significant decrease in leaf Fe and Zn concentration in the current study is as a result of a combined effect of P application, moisture regimes and genotypes. A study by Yue *et al.*, (2012) found that under favorable moisture condition, leaf concentration of Fe and Zn decrease with increasing P application in bean. The leaves of plants act as the source of assimilates which is accumulated in the grains as the sink, hence decreasing the leaf concentration due to the translocation of assimilates from the leaves to the grain (Li *et al.*, 2006). Moisture enhances the translocation of assimilates (Fe and Zn) from the leaves to the sink such as flowers, pods and grains (Uauy, 2006).

The process leading to decreased concentration of Fe and Zn in the leaves is attributed to P stimulation of growth and subsequently diluting Fe and Zn concentration or partly due to P induced Zn deficiency (Amin *et al.*, 2014). Irrespective of P application beans grown in moisture stressed conditions had higher overall leaf concentration of Fe and Zn than those grown under adequate moisture condition in the current study. This is attributed to decreased translocation of minerals such as Fe and Zn due to moisture stress and hence retention in the leaves (Uauy, 2006). In addition to retention, low yield biomass in beans grown under moisture stress conditions lead to a decreased dilution effect (Fan *et al.*, 2008; Murphy *et al.*, 2008).

5.1.4 Comparison of leaf and grain concentration of Zn and Fe in three bean genotypes.

Higher concentration of Fe and Zn in the leaves than in grains in the current study may be attributed to the role of leaves as the source of assimilates which is accumulated in grains as the sink (Li *et al.*, 2006). It follows that in a fully grown leaf, concentration of Fe and Zn is

maintained naturally higher in leaves than in grain in order for sink and source relation to exist (Visperas *et al.*, 1976).

A significant positive correlation between grain Fe and Zn concentration and leaf Fe and Zn concentration suggests that genetic factors for increasing Fe and Zn are co-segregating with genetic factors for increasing Zn (Susan and George, 2010). While a significant negative correlation between the grain yield and leaf Fe concentration and grain yield and leaf Zn concentration implies that an increase in grain yield correlates with a decrease in leaf concentration of Fe and Zn suggesting that increase in grain yield will incur a tradeoffs in leaf Fe and Zn harvest. Based on this study, improvement of soil moisture is important in order to increase the grain concentration of Fe and Zn especially in communities where human consumption of bean grain is more preferred than leaf recipe (Brian *et al.*, 2011). The current study implies that P application rate should be in consideration of grain yield as well as grain Fe and Zn harvest since the optimal P application rate for grain harvest (60kg ha^{-1}) which is higher than the optimal requirement for grain harvest of Fe and Zn under (40kg ha^{-1}).

5.2 CONCLUSIONS

- Application of P fertilizer up to 60kg ha⁻¹, maintaining adequate soil moisture throughout the growing period coupled with growing high yield bean genotypes led to increase in grain yield.
- Application of P fertilizer up to 40kg ha⁻¹, maintaining adequate soil moisture throughout the growing period and growing superior bean genotypes Awash Melka (V₂) led to an increased grain concentration of Fe and Zn.
- Beans that experienced soil moisture stress during and after flowering had higher leaf Fe and Zn concentration than those that never experienced soil moisture stress.
- Beans grown under adequate soil moisture conditions had higher grain Fe concentration than those that experienced soil moisture stress during and after flowering, however, moisture stress experienced during and after flowering had no effect on the overall grain Zn concentration.

5.3 RECOMMENDATIONS

- Farmers should be encouraged apply P fertilizer at a rate of 60kg ha^{-1} , maintain adequate soil moisture throughout the bean growing period in addition to planting superior Awash Melka (V_2) bean genotype.
- In order to have high bean grain concentration of Fe and Zn, farmers should apply P fertilizer up to 40kg ha^{-1} , maintained adequate soil moisture throughout the growing period and grow Awash Melka (V_2) bean genotype which has higher grain concentration of Fe and Zn.
- Future study should be carried out on the effect of P application and soil moisture stress experienced before the flowering period in the three bean genotypes on grain yield and leaf and grain tissue concentration of Fe and Zn in the three bean genotypes.

REFERENCES

- Abdul, J. B. K. and Saudi, H. M. (2012).** Effects of phosphorus on biological Nitrogen fixation in soya bean under irrigation using saline water By Ezra Research Station, Ezra, Daraa, Syria. *Global Journal of Science Frontier Research Agriculture & Biology*12: 1.
- Al- Farhan, H. N. and Al-Rawi, D.F. (2002).** The effect of some phosphorous fertilizers on the quality and yield of Zea mays. *Journal of Natural and Applied Science* 1: 33-40.
- Allard, R.W and Bradshaw, A.D. (1964).** Implications of genotypes-environmental interactions in applied plant breeding. *Crop Science* 4: 503-508.
- Alloway, B. J. (2004).** Zinc in Soils and Crop Nutrition. International Zinc Association (IZA) Communications. IZA publications 2: 30-64.
- Amin, S., Zaharah, A. R., Che F. I., Hanafi, M. M. and Hamed, Z. (2014).** Interaction Effects of Phosphorus and Zinc on their Uptake and P Absorption and Translocation in Sweet Corn (*Zea mays* var. *Saccharata*) Grown in a Tropical Soil. *Asian Journal of Plant Sciences*13: 129-135.
- Ann, B., Xiurong, W., Xiaolong, Y., and Hong, L.(2010)** Genetic improvement for P efficiency in soybean: a radical approach: *Annals of Botany*1: 215–222.
- Astudillo, Carolina, B. and Matthew, W. (2008).** Seed iron and Zn content and their response to P fertilization in 40 Colombian bean varieties. *Agronomia Colombiana*3: 471-476.
- Barben, S. A., Hopkins, B. G., Jolley V. D., Webb, B. L., Nichols B. A. and Buxton, E. A. (2011).** Zinc, manganese and phosphorus interrelationships and their effects on iron and copper in chelator-buffered solution grown russet burbank potato. *Journal of Plant Nutrition* 34:1144–1163.
- Beebe, R. P, M., Yan, X., Blair, W. M., Pedraza, F., Munoz, F., Lynch, P. J. (2006).** Quantitative Trait Loci for Root Architecture Traits Correlated with Phosphorus Acquisition in Common

Bean. (Genomics, Molecular Genetics & Biotechnology). *Crop Science* 46: 413-423.
<http://dx.doi.org/10.2135/cropsci2005.0226>

Blair, M. W., Giraldo M. C., Buendia H. F., Tovar E., Duque M. C. and Beebe S. E. (2006). Microsatellite marker diversity in common bean (*Phaseolus vulgaris* L.) *Theoretical and Applied Genetics* 113: 100–109.

Bouis, H. E. (2003). Micronutrient fortification of plants through plant breeding: can it improve nutrition in man at low cost. *Proceedings of nutrition society* 62:403-411.

Brevedan, R. E. and Egli, D. B. (2003). Short Periods of Water Stress during Seed Filling, Leaf Senescence, and Yield of Soybean. *Crop Science Society of America* 43:2083-2088.
<https://www.crops.org/publications/cs/abstracts/43/6/2083?access=0&view=pdf>

Brian, W., Akibode, S. and Maredia, M. (2011). Nutritional content of bean leaves. Global and regional trends in production, trade and consumption of food legume crops. Report submitted to the (CGIAR) Standing Panel on Impact Assessment.
<http://www.livestrong.com/article/454709-the-nutritional-content-of-bean-leaves/#ixzz2YA5k9zcv>

Broughton, W. J., Hernandez, G., Blair, M., Beebe, S., Gepts, P. and Vanderleyden, J. (2003). Beans (*Phaseolus* spp.). Model food legume. *Plant Soil* 252:55–128.

Buruchara, R. (2007). Background information on Common Beans (*Phaseolus vulgaris* L.) in Biotechnology, Breeding & Seed Systems for African Crops.
<http://www.africancrops.net/rockefeller/crops/beans/index.htm>.

Buruchara, R. (2006). Better beans for Africa. PABRA periodic publication. Coordinator Kawanda Agricultural. (www.ciat.cgiar.org/africapabra.htm)

Cakmak, I. and Marschner, H. (1986). Mechanism of P-induced zinc deficiency in cotton. Zinc deficiency enhanced uptake rate of P. *Physiologia Plantarum* 68: 483-490.

- Caradus, J.R.,(1994).**Selection for improved adaptation of white clover to low P and acid soils.*Euphytica* 77: 243-250.
- Carbonell, S.A.M., Azevedo,F. J.A., Dias, L.A.S., Garcia, A.A.F. and Morais, L.K. (2004).**Common Bean cultivars and line interactions with environments. *Scientia Agricola* 61:169-177.
- Cassman, D, K. G. A., & Walters, D. T. (2002).** Agroecosystems, nitrogen use efficiency, and nitrogen management. *Ambio* 31: 132-140.
- Cichy, K.A., Forster, S., Grafton, K.F. and Hosfield, G.L. (2005).** Inheritance of seed zinc accumulation in navy bean. *Crop Science* 45: 864-870.
- Cooke, G.W.(1982).** Fertilizer for Maximum yield. Granada Publishing Limited. 1: 3-59.
- Davood,H.(2013).** Phosphorus fertilizers effect on the yield and yield components of faba bean (*Vicia faba* L.) Department of Horticultural Science, Rasht Branch, Islamic Azad University, Rasht, Iran. *Annals of Biological Research* 2:181-184.
- Don, S. D.(2007).**Increasing Small Scale Farmer Productivity through Improved Inputs and Management System. Agricultural Development Initiative II.A Proposal for a Soil Health Program. AGRA publication 1:2.
- Erin L C. and Mary, L. G. (2002).**Iron stress in plants. *Genome Biology*3: 8.
- Fabricio, W. Á., Valdemar, F., Allan, K.S. L., Patrícia A. Á., Douglas, J. M., Elaine M. S, G. , Daniel K. Y. T. (2013):** Effect of Phosphite supply in nutrient solution on yield, phosphorus nutrition and enzymatic behavior in common bean (*Phaseolus vulgaris*L.) plants. *Australian Journal of Crop Science* 5:713-722.
- Fageria, N. K., Baligar, V. C. and Clark, R. B. (2006).** Physiology of Crop Production. New York: The Haworth Press 345.
- Fageria, N. K., and L. F. Stone. (2006).** Physical, chemical and biological changes in the rhizosphere and nutrient availability. *Journal of Plant Nutrition* 29: 1327–1356.

- Fageria, N. K., Baligar, V. C., Li, Y. C., (2008).** The role of nutrient efficient plants in improving crop yields in the twenty first century. *Journal of Plant Nutrition* 31:1121-1157.
- Fan, M.S., Zhao, F. J., Fairweather, S., Poulton, P., Dunham, S. and McGrath, S. (2008).** Evidence of decreasing mineral density in wheat grain over the last 160 years. *Journal of Trace Elements in Medicine and Biology* 22:315–324.
- FAO, (2010). Statistics division 2010, Food Balance Sheet
- Flora, M., Osana, B. F. and Nathan, R. (2011).** Breeding climate-proof beans to protect the "Poor Man's Protein". (CIAT bulletin). <https://ccafs.cgiar.org/blog/breeding-climate-proof-beans-protect-poor-mans-protein#.VZqA3Buqqko>
- Gathu, E. W. (2012)** Physical and chemical characterization of advanced drought tolerant bean lines for canning purposes.
URI: <http://erepository.uonbi.ac.ke:8080/xmlui/handle/123456789/7324>
- George, M. T. and Susan, N. M., (2010).** Diversity of common bean (*Phaseolus vulgaris L.*) Genotypes in iron and zinc contents under screen house conditions. *African Journal of Agricultural Research* 8: 738-747.
- Georgina, H., Mario, R., Oswaldo, V. L., Mesfin, T., Michelle, A. G., Tomasz, C., Armin, S., Maren, W., Alexander, E., Foo, C., Hank, C. W., Miguel, L., Christopher, D. T., Joachim, K., Michael, K. U., and Carroll, P. V. (2007)** Phosphorus Stress in Common Bean: Root Transcript and Metabolic Responses. *Plant Physiology* 144:751-757.
- Gianquinto, G., Abu-Rayyan, A., Tola L.D., Piccotino, D. and Pezzarossa, B. (2000).** Interaction effects of P and zinc on photosynthesis, growth and yield of dwarf bean grown in two environments. *Plant and Soil* 220: 219-228.
- Giller, K. E (2001).** Nitrogen Fixation in Tropical Cropping Systems, 2nd edn. Wallingford, United Kingdom: CAB International.

- Goff, S. A., Ricke, D., Lan, T. H., Presting, G., Wang, R., Dunn, M., Glazebrook, J., Sessions, A., Oeller, P., Varma, H. (2002).** A draft sequence of the rice genome (*Oryza sativa* L. ssp. *japonica*). *Science* 296:92-100.
- Gomez, O. (2004).** Evaluation of Nicaraguan common bean (*Phaseolus vulgaris* L.) 49 landraces. Doctoral thesis, Department of ecology and crop production sciences Uppsala, Swedish University of Agricultural Sciences.
- Graham, R. D., (1984).** Breeding for nutritional characteristics in cereals. *Advances in Plant Nutrition* 1: 57-102.
- Graham, P. H. and Ranalli, P. (1997).** Common Beans (*Phaseolus Vulgaris* L.). *Field Crops Research* 3: 131-146. <http://www.sciencedirect.com/science/article/pii/S0378429097001123>
- Graham, R., D. Senadhira, S., Beebe, C., Glesias, I., and Monasterio, I. (1999).** Breeding for micronutrient density in edible portions of staple food crops: Conventional approaches. *Field Crops Research* 60: 57-80.
- Graham, P. H., Rosas, J. C., Estevez, de J. C., Peralta, E., Tlusty, B., Acosta-Gallegos J., Arraes, P. P. A. (2003).** Addressing edaphic constraints to bean production: The bean/cowpea project in perspective. *Field Crops Research* 82: 179–192.
- Hammond, J. P., Broadley, M. R., White, P. J., King, G. J., Bowen, H. C., Hayden, R., Meacham, M. C., Mead, A., Overs, T., Spracklen, W. P., Greenwood, D. J., (2008).** Shoot yield drives phosphorus use efficiency in Brassica oleraceae and correlates with root architecture traits. *Journal of Experimental Botany* 60: 1953-1968.
- Hinga, G., Muchena, F. N. and Njihia, C. M. (1980).** Physical and Chemical Methods of Soil Analysis. National Agricultural Laboratories Manual.
- Ho, L. C. (1988).** Metabolism and compartmentation of imported sugars in sink organs in relation to sink strength. *Annual Review of Plant Physiology and Plant Molecular Biology* 39: 355-378.
-

- Ho,L.C. (1992).** Fruit growth and sink strength. In: Marshall C, Grace J, eds. Fruit and seed production: aspects of development environmentalphysiology and ecology. Society of Experimental Biology, Seminar series.*Cambridge University Press* 101-24.
- Ho,M.D.,Rosas,J.C.,Brown,K.M. and Lynch,J.P. (2005).** Root architectural tradeoffs for water and P acquisition.*FunctionalPlant Biology*32:737–748. <http://dx.doi.org/10.1071/FP05043>
- Huang, C.Y., Barker, S.J., Langridge, P., Smith, F.W. and Graham, R.D. (2009).** Zinc deficiency up-regulates expression of high affinity phosphate transporter genes in both phosphate sufficient and deficient barley (*Hordeumvulgare*L. CvWeeah) roots. *Plant Physiology*124: 415-422.
- Joe,N. A., Francis, Z., Nazim, C. and Don, F.(2013).** Evaluation of manure- derived struvite as a P source for canola. *Canadian Journal of Plant Science* 93: 419-424.
- Jaetzold, R. and Schmidt, H. (1983).**Farm management handbook of Kenya Part C.Natural conditions and Farm Management Information 2nd EditionPublished by Ministry of Agriculture, Farm Management Section. Government Printer, Nairobi, Kenya 1:17-18.
- Jerome Nriagu, (2007).** Zinc Deficiency in Human Health. *Elsevier* 1:1-8.
- Kamanga, B. C. G., Whitbread, A., Wall P., Waddington, S. R., Almekinders, C. and Giller, K. E. (2010).** Farmer evaluation of P fertilizer application to annual legumes in Chisepo, Central Malawi. *African Journal of Agricultural Research* 8: 668-680.
- Kambewa, S. P. (1997).** The Bean Sub-sector in Malawi: Historical developments, current status and policy issues. A Master of Science Thesis, Department of Agricultural economics, Michigan State University, USA.
- Karanja, D., Setegn G. E., Capitoline R., Kimani, P. M., Kweka, S. O. and Butare, L. (2011).** Value Added Bean Technologies for Enhancing Food Security, Nutrition, Income and Resilience to cope with Climate Change and Variability Challenges in Eastern Africa (ILRI).
-

- Katungi, E., Farrow A., Chianu, J., Sperling, L. and Beebe, S. (2009).** Common bean in Eastern and Southern Africa: a situation and outlook analysis. CIAT Publication.
- Kibe, J. M., Odum, H. and Macharia, P. N. (1981).** Detailed soil survey Report no D23. KARI. 29:1327-1356.
- Liao, H., Yan, X., Rubio, G., Beebe, E. S., Blair, W. M., & Lynch, P. J. (2004).** Genetic mapping of basal root gravitropism and phosphorus acquisition efficiency in common bean. *Functional Plant Biology* 31: 959-970. <http://dx.doi.org/10.1071/FP03255>
- Li, H.Y., Zhu, Y. G., Smith, S. E., Smith, F. A. (2003).** Phosphorus-zinc interactions in two barley cultivars differing in phosphorus and zinc efficiencies. *Journal of Plant Nutrition* 26: 1085-1099.
- Li, H., Huang G., Meng Q., Ma, L., Yuan, L., Wang, F., Zhang, W., Cui Z., Shen, J., Chen, X., Jiang, R. and Zhang, F. (2011).** Integrated soil and plant phosphorus management for crop and environment in China. *A review of Plant Soil* 349:157–167.
- Li, X., H. Wang, H., Li, L., Zhang, N., Teng, Q., Lin, J., Wang, T., Kuang, Z., Li, B., Li, A., Zhang and Lin, J. (2006).** Awns play a dominant role in carbohydrate production during the grain-filling stages in wheat (*Triticum aestivum* L.). *Physiologia Plantarum* 127: 701–709.
- Loggerenberg, van Magdalena. (2004).** Development and application of a small-scale procedure for the evaluation of small white beans (*Phaseolus vulgaris* L.). PhD Thesis. University of Free State, Bloemfontein, South Africa.
- Loneragan, J.K, Grove T. S., Robson, A. D. and Snowball, K. (1979).** P toxicity as a factor in zinc-P interaction in plants. *Soil Science Society of America Journal* 43: 966-972.
- Loneragan, J. K, Grunes, D.L., Welch, R. M., Aduayi, E.A, Tengah, A., Lazar, V.A. and Cary E. E. (1982).** P accumulation and toxicity in leaves in relation to zinc supply. *Soil Science Society of America Journal* 46: 345-352.
-

- Lynch, J. P. and Brown, K.M.(2001).** Topsoil foraging - an architectural adaptation of plants to low P availability. *Plant Soil* 237: 225–237.
- Lynch, J.P, Brown, K.M. (2008).** Root strategies for P acquisition. In: White P.J. and Hammond, J.P. (Eds.), *The Ecophysiology of Plant–P Interactions*. Springer.
- MacDonald, G. K., Bennett, E. M., Potter, P. A. and Ramankutty, N. (2011).** Agronomic phosphorus imbalances across the world's croplands. *Proceedings of the National Academy of Sciences* 108:3086–3091.
- Marcelis, L.F.M. (1996).** Sink strength as a determinant of dry matter partitioning in the whole plant. *Journal of Experimental Botany* 47: 1281-1291.
- Marschner, H. and Cakmak I. (1986).** Mechanism of P-induced zinc deficiency in cotton. II. Evidence for impaired shoot control of P uptake and translocation under zinc deficiency. *Physiologia Plantarum* 68: 491-496.
- Marschner, H. (1995).** Mineral nutrition of higher plants. Elsevier 5: 131-176.
- McDonald, G. K., Graham, R. D., Lloyd, J., Lewis, J., Lonergan, P., Khabaz-Saberi, H. (2001).** Breeding for improved zinc and manganese efficiency in wheat and barley. Proceeding of the 10th Australian Agronomy Conference. Department of Plant Science, Waite Institute, Glen Osmond, SA, Hobart.
- McKenzie, R. H. and Middleton, A. (1997)** Phosphorus Fertilizer Application in Crop Production. *Agriculture and Rural development* 542: 3
- Miller, C. R., Ochoa, I., Nielsen, K. L., Beck, D. and Lynch, J. P. (2003).** Genetic variation for adventitious rooting in response to low P availability: potential utility for P acquisition from stratified soils. *Functional Plant Biology* 30: 973–985.
- Ministry of Agriculture, Republic of Kenya (2011).** In: Ministry of Agriculture Annual report on production of field beans. Vol 1, MOA. Government Printer, Kenya
-

- Moraghan, J. T., Padilla, J., Etchevers, J. D., Grafton, K. and Acosta, G. J.A. (2002).** Iron accumulation in seed of common bean. *Plant and Soil*246: 175-183.
- Muasya, R. M. and Auma, E. O. (2003).** Relationship between seed quality variation and bulk seed quality in common bean (*Phaseolus vulgaris*) African Crop science Conference Proceedings.*African Crop Science Society*6: 31-37.
- Murphy, K. M., Reeves, P. G. and Jones, S. S. (2008).** Relationship between yield and mineral nutrient concentrations in historical and modern spring wheat cultivars.*Euphytica* 163:381–390.
- Njunguna, S.K., Ngegwa, A. M. M., Van Rheenen, H. A. and Mukunya, D. M. (1980).**Bean production in Kenya.In potential for field beans in Eastern Africa. Proceedings of a regional workshop held in Lilongwe, Malawi 9-14 march 1980.
- Norman, B.,(2008).**Fertilizers Role in world food Production.Bulletin. *Published by Fertilizer institute* 1: 1-4.
- Odum,E. P. (1989).** Input management of production systems. *Science* 243: 177-182.
- Pachico, D., Rachier, G. O., Kimani, P. M., Juma, R. and Ongadi, L. (1993).** The demand for bean technology, (Bean research activities at KARI Kakamega - Unpublished report).
- Pereira, H. S., Melo, L. C., Faria, L. C., Del Peloso, M. J., Costa, J.G. C., Rava, C. A. and Wendland.(2009).**A. Adaptability and stability of common bean genotypes with carioca grain type for central Brazil. *PesquisaAgropecuariaBrasileira* 44: 29-37.
- Priscila, Z. B., Bruno, R. O., Lorrana, N. N. N., Wellington, M. R. S., Helton, S. P., Cleber, M. G., Leonardo, C. M., Maria, J. D. P. (2008).** Effect of The Environment On Zinc And Iron Levels In Common Beans.Embrapa Rice and Beans, and Uni-Anhangüera-GO
<http://www.alice.cnptia.embrapa.br/alice/bitstream/doc/854603/1/BIC146.pdf>
-

- Ramalho, M.A.P., Abreu, A.F. B. and Santos, P.S.J. (1998).** Interaction genotype x seasons, years and locations in the evaluation of bean cultivars in the South and Alto Paranaíba in Minas Gerais. *Ciencia e Agrotecnologia* 22:176-181.
- Remenyik, J. and Nemeske, E. (2010).** Study of the defensive mechanism against drought in French bean (*Phaseolus vulgaris* L .) varieties. *Acta physiol plant* 32: 1125–1134.
- Rengel, Z. and Graham, R. D. (1995).** Importance of seed Zn content for wheat growth on Zn-deficient soil. *Plant Soil* 173: 259-265.
- Robson, A.D. and Pitman, M.G. (1983).** Interactions between nutrients in higher plants. In: Lauchli, A, Bielecki, R. L., eds. *Encyclopaedia of plant physiology* 5:287: 312.
- Ronno, W., Otsyula, R. and Kimani, P. (2001).** Revised list of varieties released, *CIAT Standard Methods*, Scale 1-9. Kenya Seed Company and PABRA Data 1: 60.
- Rubio, G., Liao, H., Yan, X. and Lynch, J.P. (2003).** Topsoil foraging and its role in plant competitiveness for P in common bean. *Crop Science* 43: 598–607.
- Ryan, M. H., McInerney, J. K., Record, I. R, Angus, J. F. (2008).** Zinc bioavailability in wheat grain in relation to phosphorus fertilizer, crop sequence and mycorrhizal fungi. *Journal of the Science of Food and Agriculture* 88:1208–1216.
- Sadeghzadeh, B. (2013).** A review of zinc nutrition and plant breeding; Soil Science and Plant Nutrition, Faculty of Natural and Agricultural Sciences, University of Western Australia, Crawley WA 6009, Australia. Dryland Agricultural Research Institute (DARI), Maragheh, Iran. *Journal of soil science and plant nutrition*. <http://dx.doi.org/10.4067/S0718-95162013005000072>
- SAS Institute** Incorporated. Copyright (c) 1999-2001 by Cary, N. C., USA. NOTE: SAS (r) Proprietary Software Release 8.2 (TS2M0) Licensed to Centers for Disease Control, Site 0040223003.

- Schoonhoven, A. and Voysest, O. (1991)** Common Beans: Research for Crop Improvement. *Commonwealth Agricultural Bureau International, Wallingford, UK.*
- Shah, P., Kakar, K. M., and Zada, K. (2001).** Phosphorus use efficiency of soybean as affected by phosphorus application and inoculation. *Plant Nutrition-Food Security and Sustainability of Agro-ecosystems* 670-671.
- Singh, J. P., Karamanos, R. E., Stewart, J. W. B. (1988).** The mechanism of P-induced zinc deficiency in bean (*Phaseolus vulgaris* L.). *Canadian Journal of Soil Science* 68: 345-358
- Singh, S.P. (1999).** Common Bean Improvement for the Twenty First Century, Dordrecht, Germany. Kluwer Academic Publishers 1-24.
- Siu, M.T., Saito, Maria, N. S. M., Victoria, R. L. and Reichardt, K. (1984).** The effects of N fertilizer and soil moisture on the nodulation and growth of *Phaseolus vulgaris* L. *The Journal of Agricultural Science* 103: 87-93.
- Sixbert, M. and George, M.T. (2012).** Evaluation of Common Bean (*Phaseolus vulgaris* L.) Genotypes for Adaptation to Low Phosphorus. *International Scholarly Research Network* 9: 10.
- Ssali, H. and Keya, S. O. (1986).** The effect of P and nitrogen fertilizer level on nodulation growth and dinitrogen fixation of three bean cultivars. *Tropical Agriculture* 63: 105-109/
- Susan, N. M. and George, M. T. (2010).** The Effects of the Environment on Iron and Zinc Concentrations and Performance of Common Bean (*Phaseolus vulgaris* L) genotypes. *Asian Journal of Plant Sciences* 9: 455-462.
- Szilagyi, L. (2003).** Influence of drought on seed yield components in common bean. *Bulgarian Journal of plant Physiology* 320–330.
- Tedebe, T. (2006).** Effect of common bacterial blight severity on common bean yield. *Wiley InterScience* 46:1: 41-44. <http://www.interscience.wiley.com>
-

- Uauy, C., Distelfeld, A., Fahima, T., Blechl, A., Dubcovsky, J. (2006).** A NAC gene regulating senescence, improves grain protein, zinc, and iron content in wheat. *Science* 314: 1298-1301.
- Visperas, R.M. and Venkateswarlu, B. (1976).** Source-Sink Relationships in Crop Plants. International Rice Research Institute., Los Banos, Laguna (Philippines). Plant Physiology Department.
- Vladimir, R. (2010).**The Effects Of Phosphorus Application On Soybean Plants Under Suboptimal Moisture Conditions.*Agronomy Series*53:2. http://www.revagrois.ro/PDF/2010_2_29.pdf
- Voysest, O, Valencia, M. and Amezquita, M. (1994).**Genetic diversity among Latin American Andean and Mesoamerican common bean cultivars. *Crop Science* 34:1100-1110.
- Waggoner, P. (1994).** How much land can ten billion people spare for nature?Task Force Report No. 21. Ames, Iowa, *Council for Agricultural Science and Technology*. <http://www-formal.stanford.edu/jmc/nature/nature.html>
- Wardlaw, I. F. (1990).** The control of carbon partitioning in plants. *New Phytologist* 116: 341-81.
- Webb, W. J. and Loneragan, J. F. (1988).** Effect of zinc deficiency on growth, P concentration, and P toxicity of wheat plants. *Soil Science Society of America Journal* 52 676-1680.
- Webb, W. J. and Loneragan, J. F. (1990).** Zinc translocation to wheat roots and its implications for a P/zinc interaction in wheat plants. *Journal of Plant Nutrition* 13: 1499-1512.
- Welch, R. S. and Graham, R. D. (1999).** A new paradigm for world agriculture: meeting human needs, productive, sustainable and nutritious. *Field Crops Research* 60: 1-10.
- Xavery, P., F. Ngulu, S., Kasambala., and Muthoni R., (2007).** Factors affecting Uptake of bean based technologies in Northern Tanzania, (Unpublished report).
- Xiaolong, Y., Yaoguang, L., Jonathan, L., Hong, M. A.(2008).** Adaptation of Soybean to Low Phosphorus Soils of Tropical and Subtropical South China: A Radical Approach.McKnight Foundation Collaborative Crop Research Program (CCRP), Annual report.

- Yue, Q. Z., Yan D., Ri-Yuan, C., Zhen-Ling, C., Xin-Ping, C., Russell Y., Fu-Suo, Z. and Chun-Qin, Z., (2012).**The reduction in zinc concentration of wheat grain upon increased phosphorus fertilization and its mitigation by foliar zinc application. *Plant soil* 361: 143-152.
- Zhang, Q. F. (2007)** Strategies for developing green super rice. *Proceedings of National Academy of Science* 104: 16402-16409.
- Zhu, Y. G., Smith, S. E., and Smith, F. A. (2001).** Zinc (Zn)-phosphorus (P) Interactions in Two Cultivars of Spring Wheat (*Triticum aestivum* L.) Differing in P Uptake Efficiency. *Annals of Botany* 88: 941: 945.
- Zhu, J.M., Kaepler, S.M. and Lynch, J.P., (2005).** Topsoil foraging and P acquisition efficiency in maize (*Zea mays*). *Functional Plant Biology* 32: 749–762.
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APPENDICES

Appendix 1: ANOVA Table for Interaction of P Application rates, Moisture regimes and Genotypes on Bean Grain Yield (Kg Ha⁻¹).

Dependent Variable: Grain Yield.					
Source	DF	Squares	Mean Square	F Value	Pr> F
Model		31	75244118.71	2427229.64	92.97 <.0001
Error		58	1514291.47	26108.47	
Corrected Total	89	76758410.18			
	R-Square	CoeffVar	Root MSE	Mean grain yield	
	0.980272	6.584039	161.5812	2454.134	
Source	DF	Anova SS	Mean Square	F Value	Pr> F
Stress	1	58750165.05	58750165.05	2250.23	<.0001
P rate	4	11619881.84	2904970.46	111.27	<.0001
Genotypes	2	952245.34	476122.67	18.24	<.0001
Stress x P rate	4	1394745.89	348686.47	13.36	<.0001
Stress X Genotype	2	1187117.77	593558.89	22.73	<.0001
Stress x P rate X Genotypes	16	1330971.00	83185.69	3.19	0.0006

Appendix 2: ANOVA Table for Interaction of P Application rates, Moisture regimes and Genotypes on Bean Grain Fe Concentration (Mg Kg⁻¹)

Dependent Variable: Grain Fe concentration.

Source	DF	Sum of Squares	Mean Square	F Value	Pr> F
Model	31	7789.544444	251.275627	125.37	<.0001
Error	58	116.244444	2.004215		
Corrected Total	89	7905.788889			

R-Square	CoeffVar	Root MSE	Mean grain Fe
0.985296	1.898008	1.415703	74.58889

Source	DF	Anova SS	Mean Square	F Value	Pr> F
Stress	1	208.544444	208.544444	104.05	<.0001
P rate	4	1118.511111	279.627778	139.52	<.0001
Genotypes	2	6102.755556	3051.377778	1522.48	<.0001
Stress X P rate	4	45.844444	11.461111	5.72	.0006
Stress X Genotypes	2	64.355556	32.177778	16.06	<.0001
Stress X P rate X Genotypes	16	243.777778	15.236111	7.60	<.0001

Appendix 3: ANOVA Table for the Interaction of P Application rates, Moisture regimes and Genotypes on Bean Grain Zn Concentration (Mg Kg⁻¹)

Dependent Variable: Grain Zn concentration.					
Source	DF	Sum of Squares	Mean Square	F Value	Pr> F
Model	31	454.2217778	14.6523154	39.25	<.0001
Error	58	21.6511111	0.3732950		
Corrected Total	89	475.8728889			
R-Square		<u>CoeffVar</u>	<u>RootMS</u>	<u>Mean grain Zn</u>	
	0.954502	2.728666	0.610979	22.39111	
Source	DF	Anova SS	Mean Square	F Value	Pr> F
Stress	1	0.0017778	0.0017778	0.00	0.9452
P rate	4	80.2240000	20.0560000	53.73	<.0001
Genotypes	2	324.4115556	162.2057778	434.52	<.0001
Stress X P rate	4	8.5582222	2.1395556	5.73	0.0006
Stress X Genotype	2	0.3795556	0.1897778	0.51	0.6041
Stress X P rate X Genotype	16	39.8244444	2.4890278	6.67	<.0001

Appendix 4: ANOVA Table for Interaction of P Application rates, Moisture regimes and Genotypes on Bean Leaf Fe Concentration (Mg Kg⁻¹)

Dependent Variable: Leaf Fe					
Sum of					
Source	DF	Squares	Mean Square	F Value	Pr> F
Model	31	135281.1333	4363.9075	502.93	<.0001
Error	58	503.2667	8.6770		
Corrected Total	89	135784.4000			
	R-Square	CoeffVar	Root MSE	Mean leaf Fe	
	0.996294	1.078213	2.945677	273.2000	
Source	DF	Anova SS	Mean Square	F Value	Pr> F
Stress	1	1960.00000	1960.00000	225.88	<.0001
P rate	4	65820.17778	16455.04444	1896.40	<.0001
Genotype	2	22786.06667	11393.03333	1313.01	<.0001
Stress X P rate	4	494.22222	123.55556	14.24	<.0001
Stress X Genotype	2	32718.06667	16359.03333	1885.33	<.0001
Stress X P rate X Genotypes	16	11489.20000	718.07500	82.76	<.0001

Appendix 5: ANOVA Table for Interaction of P Application rates, Moisture regimes and Genotypes on Bean Leaf Zn Concentration (Mg Kg⁻¹).

Dependent Variable: Leaf Zn Concentration.						
Sum of						
Source	DF	Squares	Mean Square	F Value	Pr> F	
Model	31	394.9344444	12.7398208	13.12	<.0001	
Error	58	56.3251111	0.9711226			
Corrected Total	89	451.2595556				
	R-Square	CoeffVar	Root MSE	Mean Leaf Zn		
	0.875182	3.476988	0.985456	28.34222		
Source	DF	Anova SS	Mean Square	F Value	Pr> F	
Stress	1	51.0760000	51.0760000	52.59	<.0001	
P rate	4	224.3984444	56.0996111	57.77	<.0001	
Genotype	2	17.4648889	8.7324444	8.99	0.0004	
Stress X P rate	4	14.7428889	3.6857222	3.80	0.0083	
Stress X Genotype	2	37.8560000	18.9280000	19.49	<.0001	
Stress X P rate X Genotype	16	45.7946667	2.8621667	2.95	0.0013	

Appendix 6: ANOVA Table for Interaction of P Application rates, Moisture regimes and Genotypes on Percentage (%) of Bean Leaf Fe Accumulated to Grain.

Dependent Variable: Percentage Leaf Fe Accumulated to Grain.					
Source	Sum of DF	Squares	Mean Square	F Value	Pr> F
Model	31	2732.979227	88.160620	141.49	<.0001
Error	58	36.139813	0.623100		
Corrected Total	89	2769.119040			
	R-Square	CoeffVar	Root MSE	mean %leaf Fe:grain	
	0.986949	2.824009	0.789367	27.95200	
Source	DF	Anova SS	Mean Square	F Value	Pr> F
Stress	1	63.369671	63.369671	101.70	<.0001
P rate	4	1055.847884	263.961971	423.63	<.0001
Genotype	2	1167.462587	583.731293	936.82	<.0001
Stress X P rate	4	10.493129	2.623282	4.21	0.0046
Stress X Genotype	2	302.059742	151.029871	242.38	<.0001
Stress X P rate X Genotype	16	133.227093	8.326693	13.36	<.0001

Appendix7: ANOVA Table for Interaction of P Application rates, Moisture regimes and Genotypes on Percentage (%) of Bean Leaf Zn Accumulated to Grain.

Dependent Variable: Percentage Leaf Zn Accumulated to Grain.

Source	Sum of DF	Squares	Mean Square	F Value	Pr> F
Model	31	10454.63744	337.24637	34.06	<.0001
Error	58	574.20978	9.90017		
Corrected Total	89	11028.84722			
R-Square	CoeffVar	Root MSE	Mean % leaf Zn: grain		
0.947936	3.958080	3.146453	79.49444		

Source	DF	Anova SS	Mean Square	F Value	Pr> F
Stress	1	503.626778	503.626778	50.87	<.0001
P rate	4	3461.136667	865.284167	87.40	<.0001
Genotype	2	5005.953556	2502.976778	252.82	<.0001
Stress X P rate	4	369.343778	92.335944	9.33	<.0001
Stress X Genotype	2	401.113556	200.556778	20.26	<.0001
Stress X P rate X Genotype	16	658.566222	41.160389	4.16	<.0001

Appendix 8: Soil Analysis Results at KARI - Katumani Experimental Plot at the Beginning of the Experiment.

Soil Properties	Content
Soil pH (water 1 : 2.5)	6.36
Total N (%)	0.11
Organic Carbon (%)	0.89
P (mg kg ⁻¹)	15
Potassium (mg kg ⁻¹)	1.14
Magnesium (mg kg ⁻¹)	2.87
Calcium (mg kg ⁻¹)	3.40
Sodium (mg kg ⁻¹)	0.16
Copper (µgg ⁻¹)	4.63
Iron (µgg ⁻¹)	15.10
Zinc (µgg ⁻¹)	6.91
Manganese (µgg ⁻¹)	0.87

Appendix 9: Field Data.

moisture stress	Repeat Rep	P rate (Kg ha ⁻¹)	genotype v\$	Grain weight (Kg ha ⁻¹)	Grain Fe (mg kg ⁻¹)	Grain Zn (mg kg ⁻¹)	Leaf Fe (mg kg ⁻¹)	Leaf Zn (mg kg ⁻¹)	Grain/leaf Fe (%)	Grain/leaf Zn %
1	1	0	V ₁	2005.3	60	19.6	291	31.5	21	62.2
1	2	0	V ₁	2488.9	63	18.9	291	31.5	22	60.0
1	3	0	V ₁	2232.9	61	18.2	291	31.5	21	57.8
1	1	20	V ₁	2631.1	69	19.6	278	29.2	24.8	67.1
1	2	20	V ₁	3100.4	69	19.3	278	29.3	24.8	65.9
1	3	20	V ₁	2872.8	69	20.0	278	29.0	24.8	69
1	1	40	V ₁	3527.1	70	20.4	240	27.2	29.2	75
1	2	40	V ₁	3640.8	74	19.0	237	27.2	31.2	69.9
1	3	40	V ₁	3441.7	72	20.1	239	27.2	30.1	73.9
1	1	60	V ₁	3868.4	67	20.4	242	28.3	26	72.1
1	2	60	V ₁	4159.9	66	20.1	242	28.3	25.6	71
1	3	60	V ₁	4081.7	64	20.3	242	28.3	26.4	71.7
1	1	80	V ₁	4088.8	64	19.7	241	25.2	26.6	78.2
1	2	80	V ₁	4181.3	62	20.0	241	28.7	25.7	69.7
1	3	80	V ₁	3925.3	63	19.1	241	29.3	26.1	65.2
1	1	0	V ₂	2719.2	81	21.0	316	30.4	25.6	69.1
1	2	0	V ₂	2717.8	80	22.1	316	29.0	25.3	76.1
1	3	0	V ₂	2887.1	81	23.6	316	31.7	25.6	74.4
1	1	20	V ₂	3393.4	85	24.6	285	28.4	29.8	86.6
1	2	20	V ₂	3143.1	85	23.6	285	28.2	29.8	83.7
1	3	20	V ₂	2929.7	85	24.6	285	28.3	29.8	86.9
1	1	40	V ₂	3413.28	92	25.6	265	26.5	34.7	96.6
1	2	40	V ₂	3697.7	90	25.5	270	26.5	33.3	96.6

1	3	40	V ₂	3612.4	95	25.3	265	26.5	35.8	95.5
1	1	60	V ₂	3868.4	90	24.7	262	23.5	34.4	100
1	2	60	V ₂	3896.8	92	25.1	262	24.8	35.1	100
1	3	60	V ₂	3854.2	88	23.4	262	22.2	33.6	100
1	1	80	V ₂	4081.7	83	23.0	261	23.3	31.8	100
1	2	80	V ₂	3839.9	83	23.8	260	23.3	31.9	100
1	3	80	V ₂	3939.5	83	22.2	261	23.3	31.8	95
1	1	0	V ₃	2545.7	70	20.2	335	30.7	20.9	65.8
1	2	0	V ₃	2446.2	73	19.7	332	29.7	22	66.3
1	3	0	V ₃	2659.5	71	20.8	330	28.7	21.5	72.5
1	1	20	V ₃	2531.5	76	24.7	309	30.0	24.6	82.3
1	2	20	V ₃	2897	74	24.9	305	30.3	24.3	82.2
1	3	20	V ₃	2801.7	75	24.6	301	27.5	24.9	89.5
1	1	40	V ₃	2659.5	80	25.5	244	26.8	31.9	95.1
1	2	40	V ₃	2986.6	83	24.8	244	26.5	35.2	93.6
1	3	40	V ₃	3150.2	85	25.2	245	26.8	34.7	94
1	1	60	V ₃	3527.1	81	24.6	236	27.0	34.3	91
1	2	60	V ₃	3100.4	79	23.7	231	24.8	34.2	95.6
1	3	60	V ₃	3327.9	84	24.7	233	26.2	36.1	94.3
1	1	80	V ₃	3640.8	71	24.4	232	26.3	34.9	92.8
1	2	80	V ₃	3128.8	72	23.2	232	26.3	31	88.2
1	3	80	V ₃	3150.2	73	24.0	232	26.3	31.5	91.3
2	1	0	V ₁	1224.5	57	18.9	341	30.0	16.7	63
2	2	0	V ₁	1167.6	56	19.4	341	30.0	16.4	64.7
2	3	0	V ₁	1123.5	58	19.0	341	30.0	17	63.3
2	1	20	V ₁	1499	64	19.6	288	29.0	22.2	67.6
2	2	20	V ₁	1483.4	64	19.8	288	29.0	22.2	68.3
2	3	20	V ₁	1450.6	63	19.5	288	29.0	21.8	67.2

2	1	40	V ₁	1523.2	67	21.0	246	29.0	27.2	72.4
2	2	40	V ₁	1792	66	20.7	243	28.9	27.2	71.6
2	3	40	V ₁	1493.3	66	20.9	239	28.7	27.6	72.8
2	1	60	V ₁	1848.9	64	19.6	269	28.0	23.58	70
2	2	60	V ₁	1593	65	19.9	252	28.5	25.8	69.8
2	3	60	V ₁	1920	65	20.3	252	28.3	25.8	71.7
2	1	80	V ₁	1962.6	59	19.4	250	24.2	23.6	80.2
2	2	80	V ₁	1807.6	59	19.3	250	28.5	23.6	67.7
2	3	80	V ₁	1706.6	57	19.2	250	26.0	22.8	73.8
2	1	0	V ₂	1564.4	77	24.4	256	30.5	30.1	80
2	2	0	V ₂	1331.2	78	23.0	256	30.5	30.5	75.4
2	3	0	V ₂	1066.7	79	21.6	256	30.5	30.9	70.8
2	1	20	V ₂	1578.6	77	24.0	242	29.7	31.8	80.8
2	2	20	V ₂	1479.1	80	23.6	242	29.7	33.1	79.5
2	3	20	V ₂	1536	79	23.8	242	29.7	32.6	80.1
2	1	40	V ₂	1720.9	85	24.2	224	28.7	37.9	84.3
2	2	40	V ₂	1806.2	90	24.2	229	28.7	39.3	84.3
2	3	40	V ₂	1766.4	87	24.2	229	28.7	38	84.3
2	1	60	V ₂	1784.9	83	24.4	230	25.7	36.1	94.9
2	2	60	V ₂	1910	84	24.4	230	30.2	36.5	80.8
2	3	60	V ₂	1955.5	81	24.4	230	29.1	35.2	83.8
2	1	80	V ₂	1863.1	81	23.8	265	28.0	30.6	85
2	2	80	V ₂	1976.9	82	22.4	256	28.0	32	80
2	3	80	V ₂	1955.5	79	22.9	261	28.0	30.3	81.8
2	1	0	V ₃	1365.3	70	22.1	370	33.3	18.9	66.4
2	2	0	V ₃	995.5	70	23.2	370	33.3	18.9	69.7
2	3	0	V ₃	1402.3	70	21.0	370	33.3	18.9	63.1

2	1	20	V ₃	1742.2	73	25.7	352	30.0	20.7	85.7
2	2	20	V ₃	1507.5	74	26.0	352	30.8	21	84.4
2	3	20	V ₃	1479.1	75	25.4	352	29.2	21.3	87
2	1	40	V ₃	1848.9	79	24.9	284	29.5	27.8	84.4
2	2	40	V ₃	1664	81	26.8	304	31.5	26.6	85.1
2	3	40	V ₃	1692.4	77	25.9	294	27.7	26.2	93.5
2	1	60	V ₃	1991.1	79	23.4	302	27.5	26.2	85.1
2	2	60	V ₃	1706.6	77	23.5	302	29.6	25.5	79.4
2	3	60	V ₃	1934.2	80	23.2	302	30.5	26.5	76.1
2	1	80	V ₃	1991.1	77	22.0	286	26.7	26.9	82.4
2	2	80	V ₃	1937	77	21.5	286	26.8	26.9	80.2
2	3	80	V ₃	1930	77	21.0	284	26.8	27.1	78.4